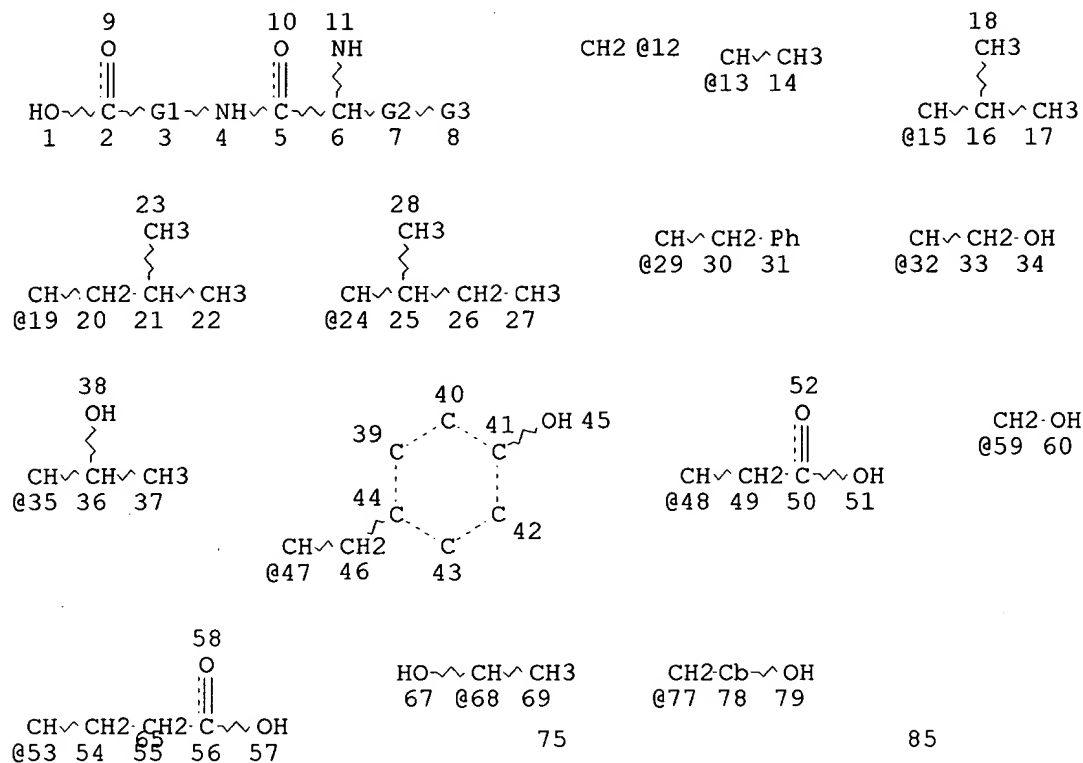


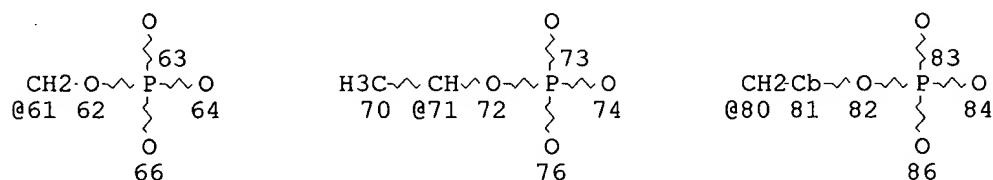
=> d que

L1

STR



Page 1-A



Page 2-A

VAR G1=12/13/15/19/24/29/32/35/47/48/53

REP G2=(0-10) CH2

VAR G3=59/61/68/77/71/80

NODE ATTRIBUTES:

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CONNECT IS E1 RC AT 65

CONNECT IS E1 RC AT 66

CONNECT IS E1 RC AT 74

CONNECT IS E1 RC AT 75

CONNECT IS E1 RC AT 76

CONNECT IS E1 RC AT 84

CONNECT IS E1 RC AT 85

CONNECT IS E1 RC AT 86

DEFAULT MLEVEL IS ATOM

GGCAT IS MCY UNS AT 78

GGCAT IS MCY UNS AT 81

DEFAULT ECLEVEL IS LIMITED

ECOUNT IS E6 C AT 78

ECOUNT IS E6 C AT 81

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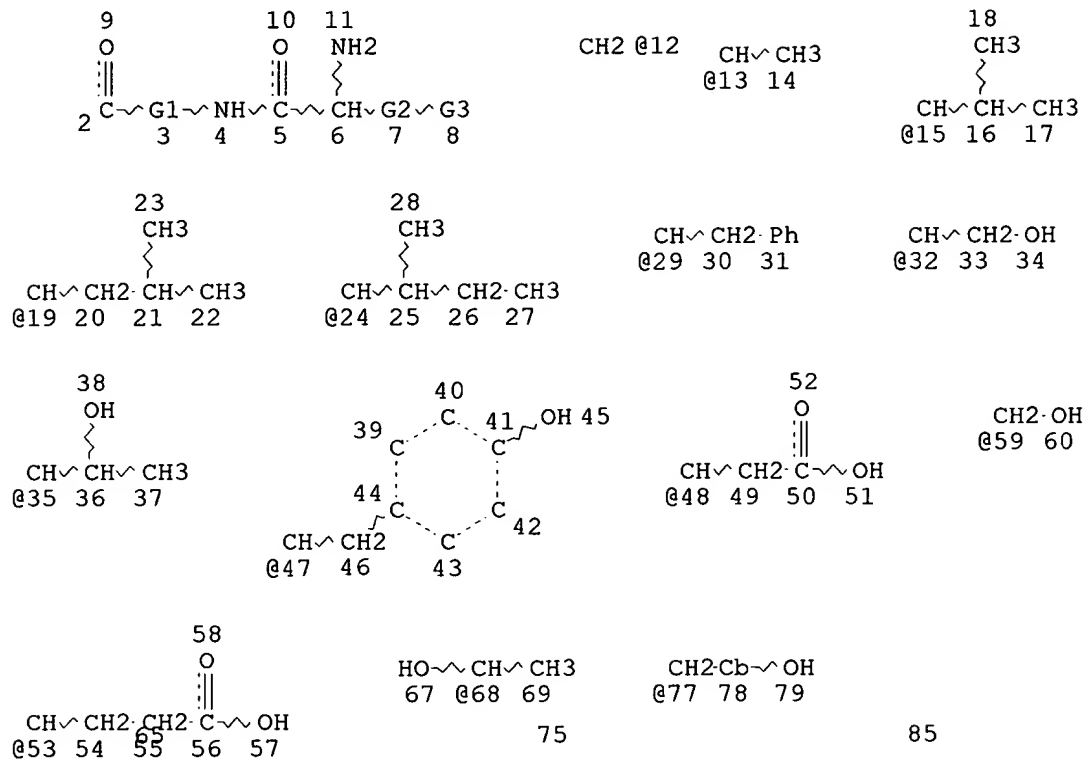
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NUMBER OF NODES IS 86

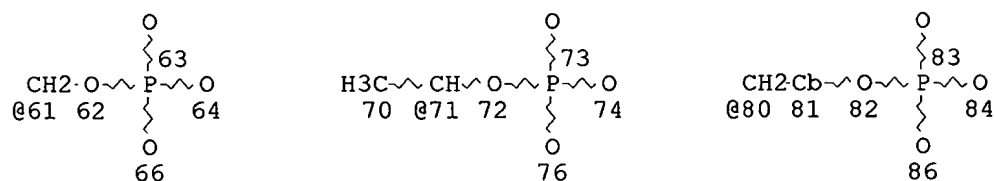
STEREO ATTRIBUTES: NONE

L3

STR



Page 1-A



Page 2-A

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VAR G3=59/61/68/77/71/80

NODE ATTRIBUTES:

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CONNECT IS E1 RC AT 65

CONNECT IS E1 RC AT 66

CONNECT IS E1 RC AT 74

CONNECT IS E1 RC AT 75

CONNECT IS E1 RC AT 76
 CONNECT IS E1 RC AT 84
 CONNECT IS E1 RC AT 85
 CONNECT IS E1 RC AT 86
 DEFAULT MLEVEL IS ATOM
 GGCAT IS MCY UNS AT 78
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 DEFAULT ECLEVEL IS LIMITED
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RSPEC 39

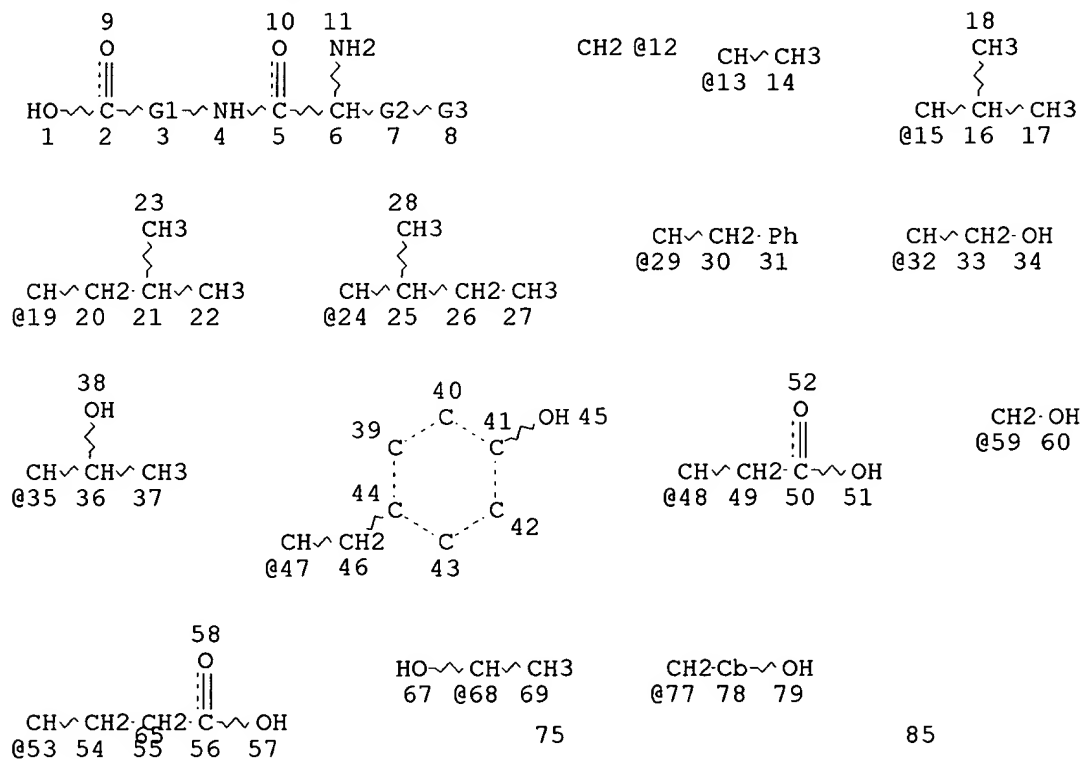
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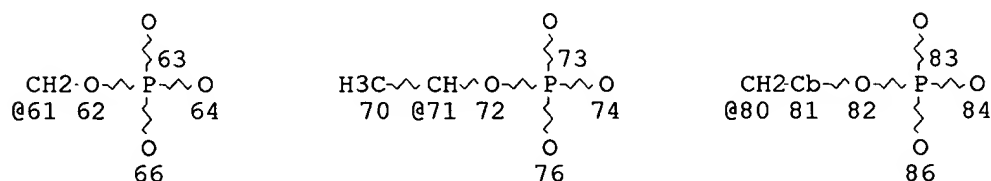
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L6 58 SEA FILE=REGISTRY SUB=L4 SSS FUL L1 AND L3

L14 STR



Page 1-A



Page 2-A

VAR G1=12/13/15/19/24/29/32/35/47/48/53

REP G2=(0-10) CH2

VAR G3=59/61/68/77/71/80

NODE ATTRIBUTES:

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CONNECT IS E1 RC AT 65

CONNECT IS E1 RC AT 66

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CONNECT IS E1 RC AT 75

CONNECT IS E1 RC AT 76

CONNECT IS E1 RC AT 84

CONNECT IS E1 RC AT 85

CONNECT IS E1 RC AT 86

DEFAULT MLEVEL IS ATOM

GGCAT IS MCY UNS AT 78

GGCAT IS MCY UNS AT 81

DEFAULT ECLEVEL IS LIMITED

ECOUNT IS E6 C AT 78

ECOUNT IS E6 C AT 81

GRAPH ATTRIBUTES:

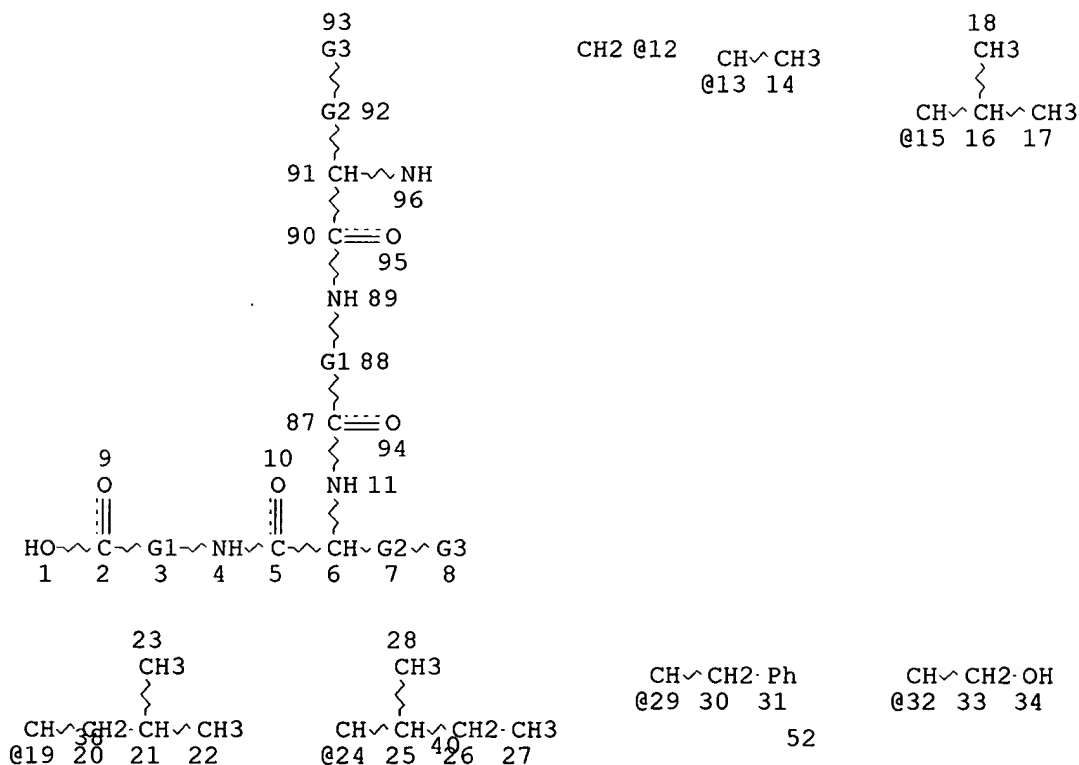
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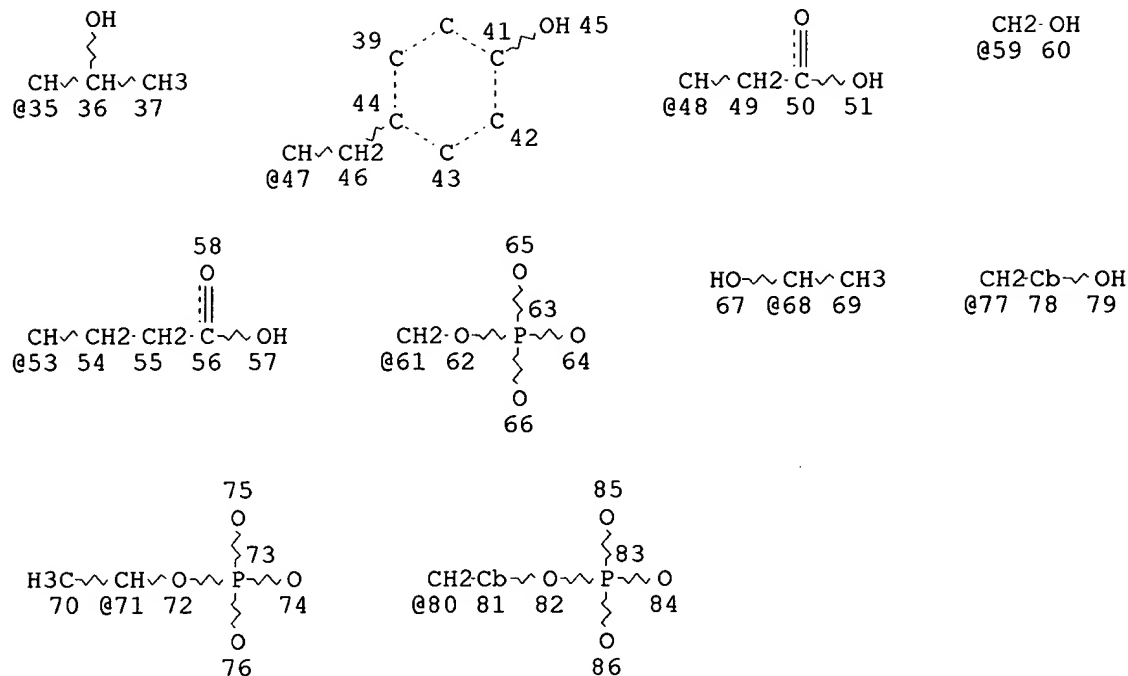
NUMBER OF NODES IS 86

STEREO ATTRIBUTES: NONE

L15 19 SEA FILE=REGISTRY SUB=L6 SSS FUL L14

L16 STR





Page 2-A

VAR G1=12/13/15/19/24/29/32/35/47/48/53

REP G2=(0-10) CH2

VAR G3=59/61/68/77/71/80

NODE ATTRIBUTES:

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 CONNECT IS E1 RC AT 65
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 CONNECT IS E1 RC AT 86

DEFAULT MLEVEL IS ATOM

GGCAT IS MCY UNS AT 78

GGCAT IS MCY UNS AT 81

DEFAULT ECLEVEL IS LIMITED

ECOUNT IS E6 C AT 78

ECOUNT IS E6 C AT 81

GRAPH ATTRIBUTES:

RSPEC 39

NUMBER OF NODES IS 96

STEREO ATTRIBUTES: NONE

L17 10 SEA FILE=REGISTRY SUB=L6 SSS FUL L16

L18 29 SEA FILE=REGISTRY ABB=ON PLU=ON L15 OR L17

L19 24 SEA FILE=HCAPLUS ABB=ON PLU=ON L18

=> d ibib ab hitstr 119 1-24

L19 ANSWER 1 OF 24 HCAPLUS COPYRIGHT 2002 ACS
 ACCESSION NUMBER: 2001:772123 HCAPLUS
 DOCUMENT NUMBER: 135:313632
 TITLE: Phosphopeptides and methods of treating bone diseases
 INVENTOR(S): Kumagai, Yoshinari; Otaka, Akira
 PATENT ASSIGNEE(S): Big Bear Bio, Inc., USA
 SOURCE: U.S., 16 pp., Cont.-in-part of U.S. 5,837,674.
 CODEN: USXXAM
 DOCUMENT TYPE: Patent
 LANGUAGE: English
 FAMILY ACC. NUM. COUNT: 2
 PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
US 6306822	B1	20011023	US 1998-193214	19981116
US 5837674	A	19981117	US 1996-675031	19960703
CA 2258661	AA	19980108	CA 1997-2258661	19970630

PRIORITY APPLN. INFO.: US 1996-675031 A2 19960703

AB Phosphopeptides which significantly reduce bone loss or weakening are provided by the invention. Also provided is a method for treating or preventing any condition assocd. with bone loss or weakening by administering the phosphopeptides by oral or injectable means.

IT 201660-41-5 201660-42-6 201660-43-7
 328896-68-0 328896-69-1

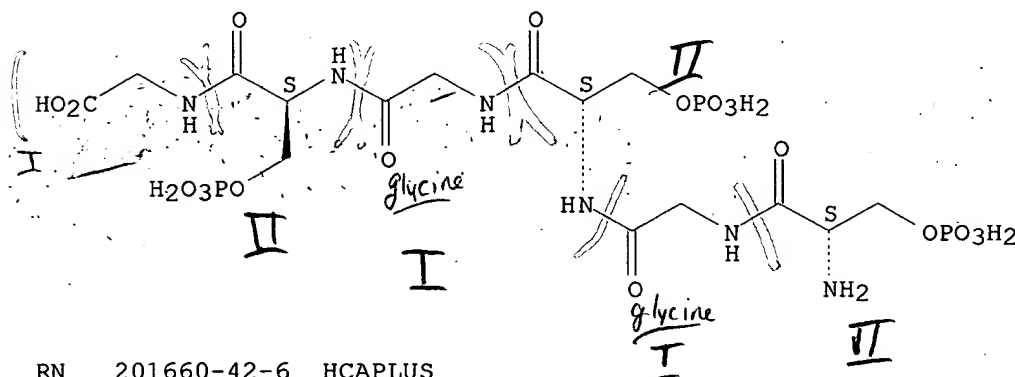
RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); THU (Therapeutic use); BIOL (Biological study); USES (Uses)

(phosphopeptides and use in bone disease treatment)

RN 201660-41-5 HCAPLUS

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Absolute stereochemistry.

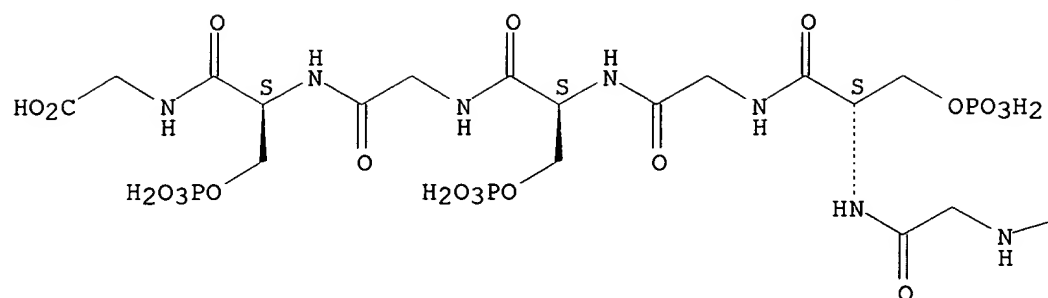


RN 201660-42-6 HCAPLUS

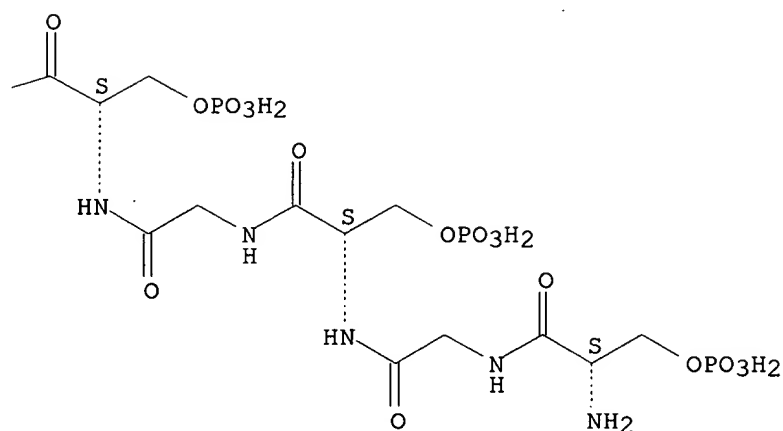
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Absolute stereochemistry.

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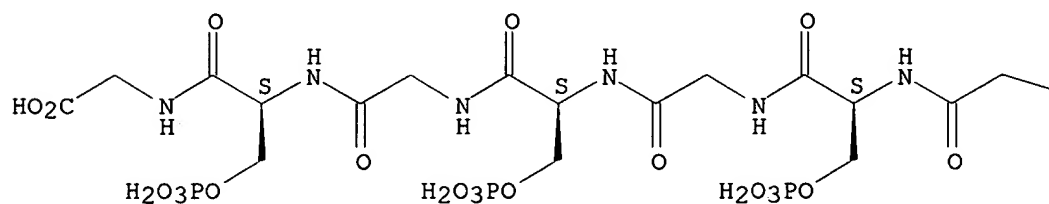


RN 201660-43-7 HCAPLUS

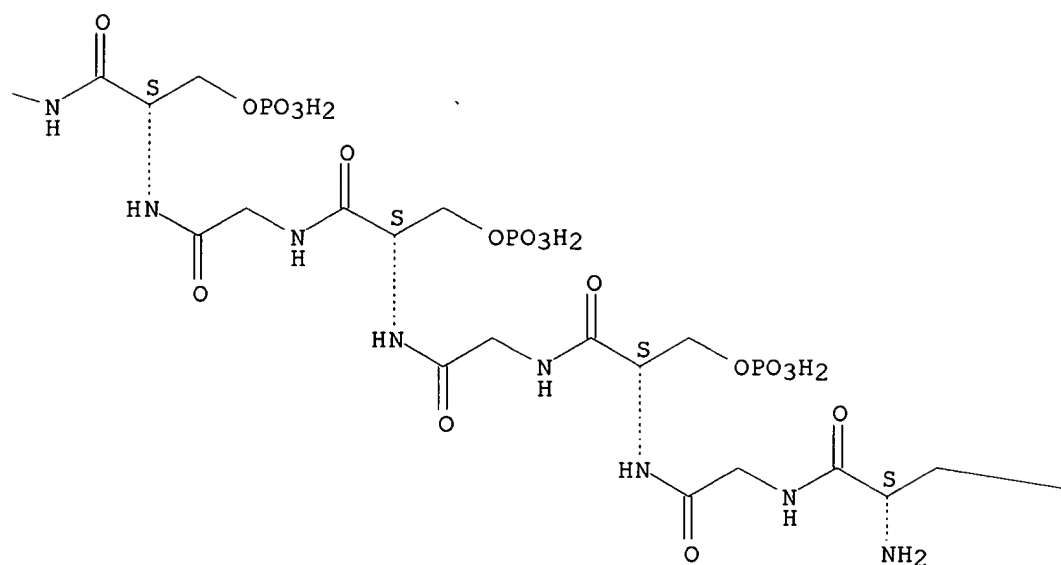
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Absolute stereochemistry.

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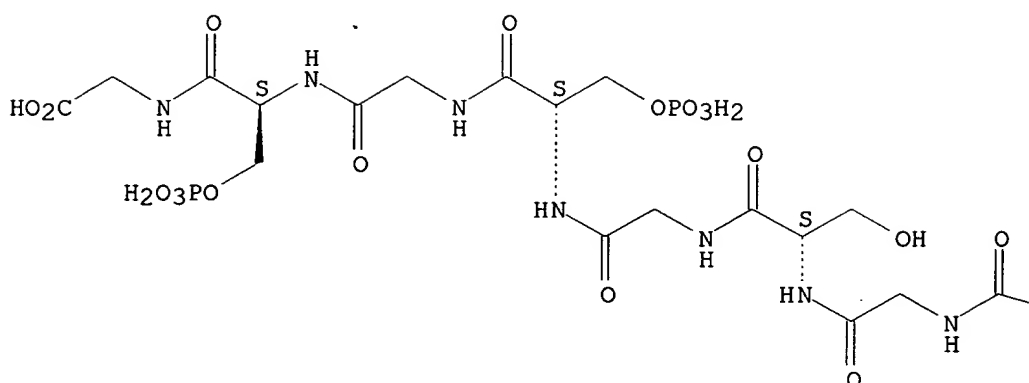
—OPO₃H₂

RN 328896-68-0 HCAPLUS

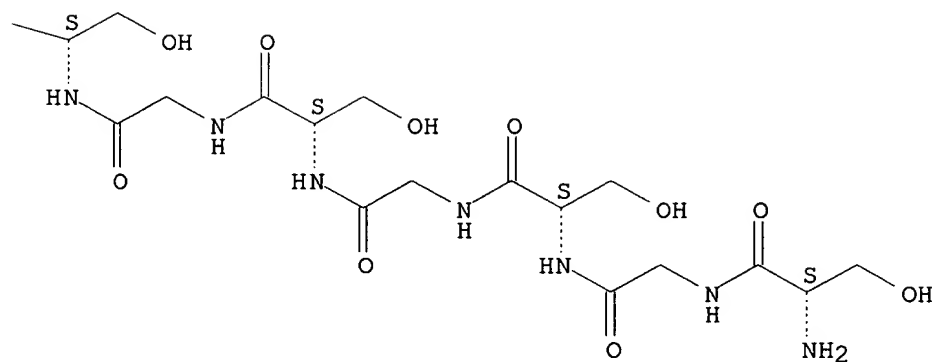
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Absolute stereochemistry.

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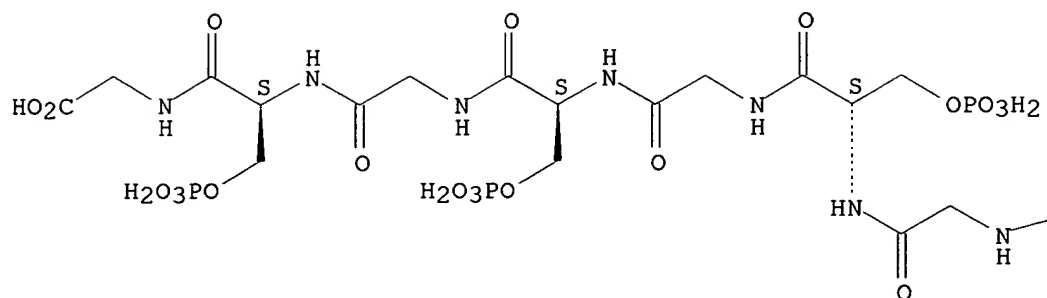


RN 328896-69-1 HCAPLUS

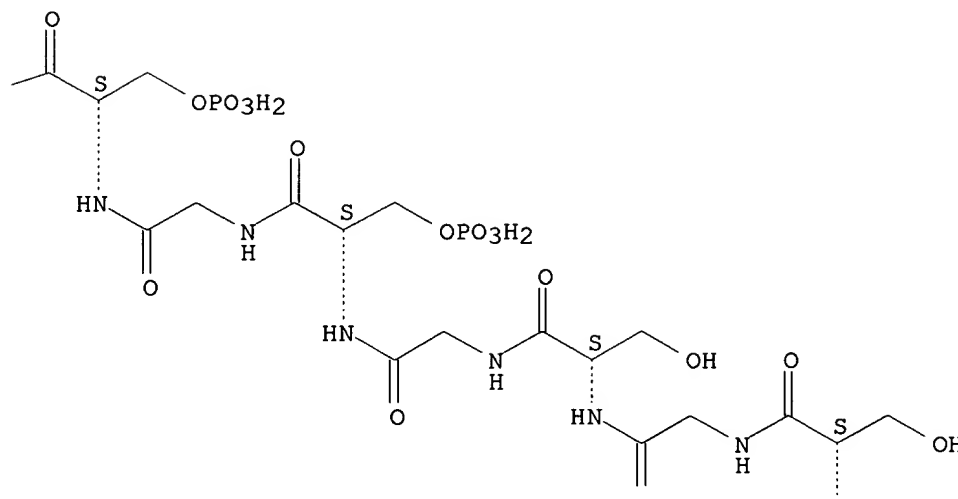
CN Glycine, L-serylglycyl-L-serylglycyl-O-phosphono-L-serylglycyl-O-phosphono-L-serylglycyl-O-phosphono-L-serylglycyl-O-phosphono-L-serylglycyl-O-phosphono-L-seryl- (9CI) (CA INDEX NAME)

Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



PAGE 2-B



REFERENCE COUNT: 14 THERE ARE 14 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L19 ANSWER 2 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 2001:167827 HCAPLUS

DOCUMENT NUMBER: 134:202705

TITLE: Methods and compositions using peptidic compounds for reducing serum phosphate levels

INVENTOR(S): Kumaqai, Yoshinari; Otaka, Akira
 PATENT ASSIGNEE(S): Big Bear Bio Inc., USA
 SOURCE: PCT Int. Appl., 37 pp.
 CODEN: PIXXD2
 DOCUMENT TYPE: Patent
 LANGUAGE: English
 FAMILY ACC. NUM. COUNT: 1
 PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 2001015720	A1	20010308	WO 2000-US22910	20000817
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG EP 1207896 A1 20020529 EP 2000-957624 20000817 R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL PRIORITY APPLN. INFO.: US 1999-152046P P 19990902 WO 2000-US22910 W 20000817				

OTHER SOURCE(S): MARPAT 134:202705

AB Peptidic compds. are provided which significantly reduce serum phosphate levels, and which, in addn., reduce bone loss or weakening. Also provided is a method for treating or preventing any condition assocd. with elevated serum phosphate levels by administering the peptidic compds. by oral or injectable means.

IT **201660-43-7 328896-68-0 328896-69-1**

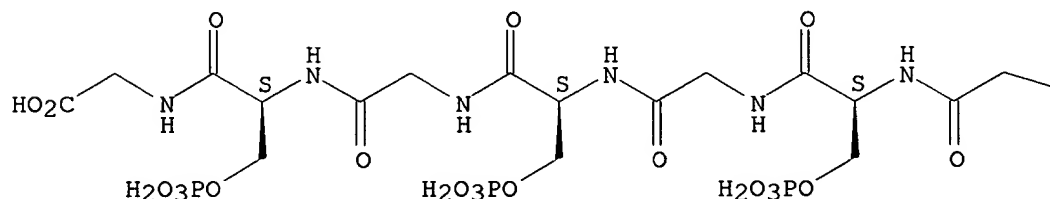
RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); PRP (Properties); THU (Therapeutic use); BIOL (Biological study); USES (Uses)
 (peptidic compds. for reducing serum phosphate levels and reducing bone loss and weakening)

RN 201660-43-7 HCAPLUS

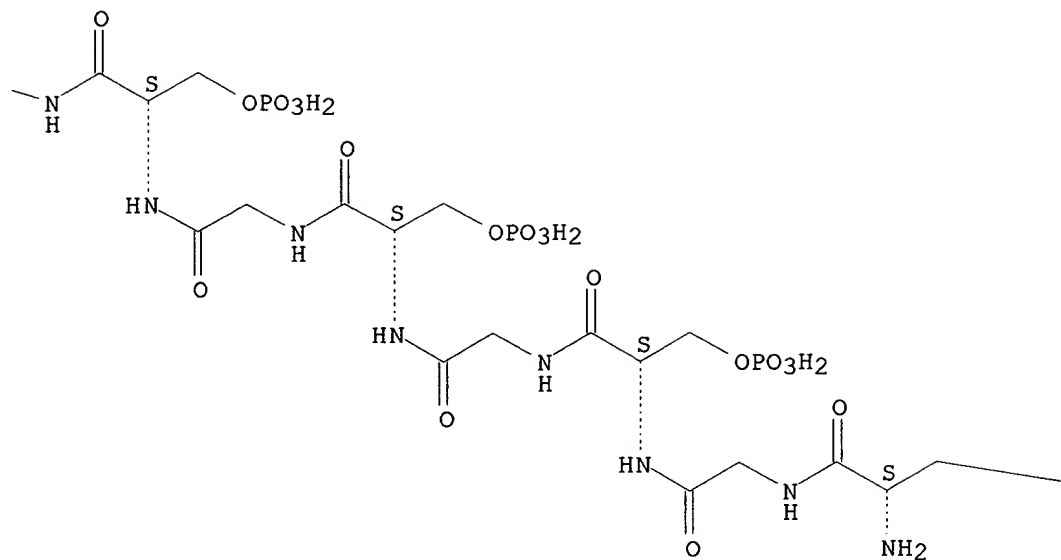
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Absolute stereochemistry.

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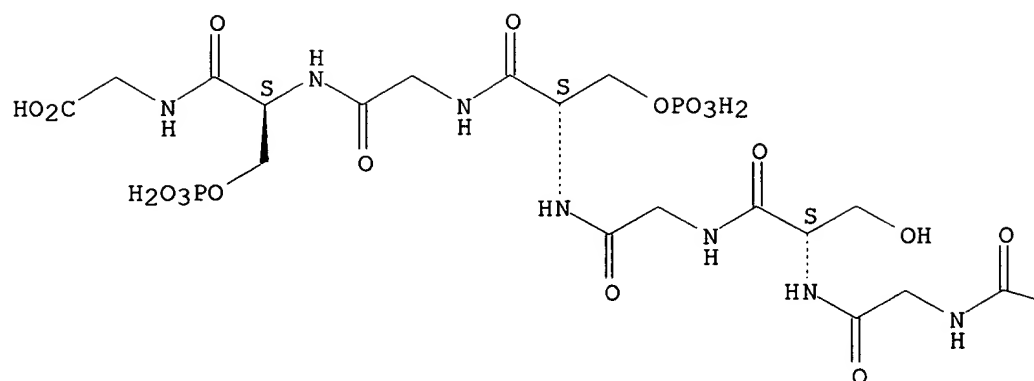
—OPO₃H₂

RN 328896-68-0 HCAPLUS

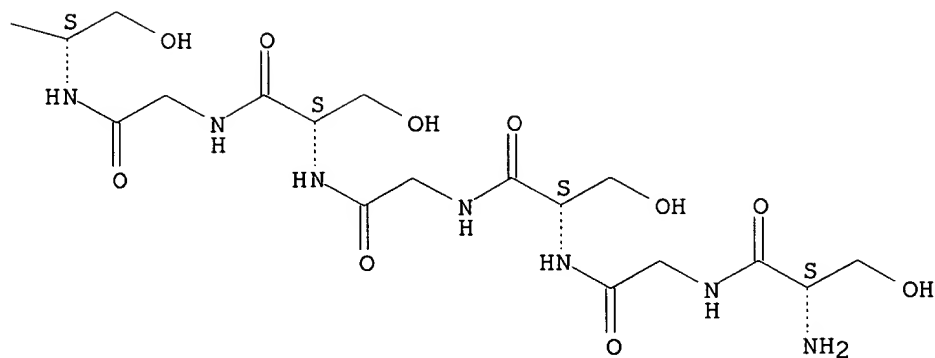
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Absolute stereochemistry.

PAGE 1-A



PAGE 1-B

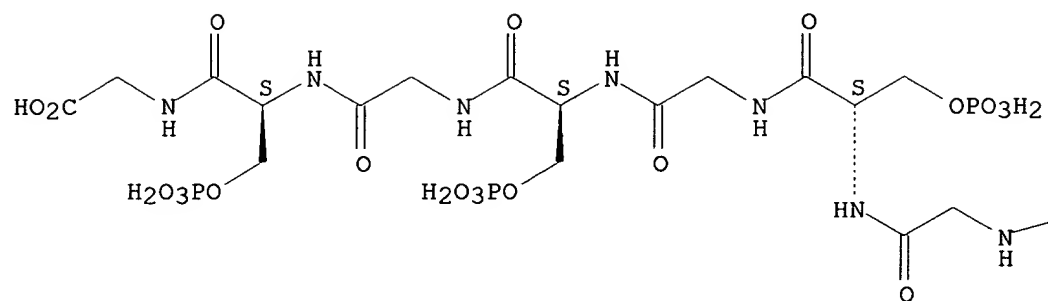


RN 328896-69-1 HCAPLUS

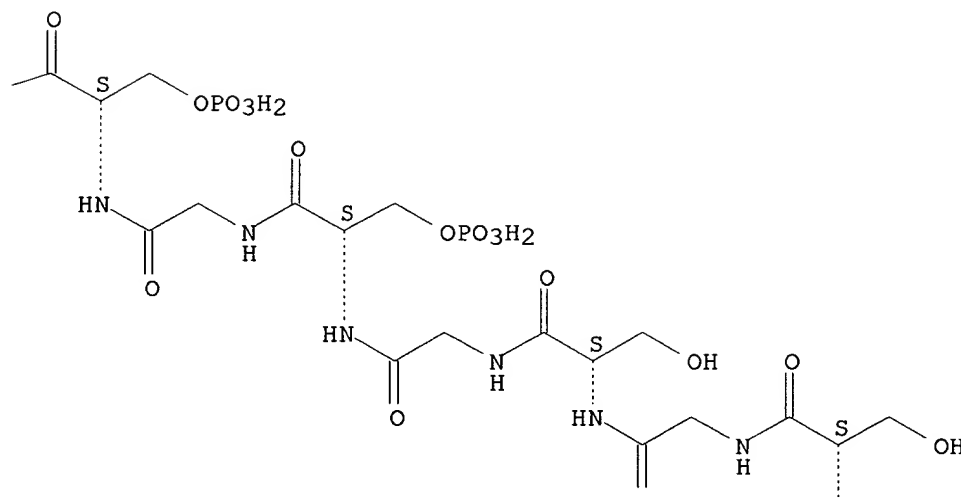
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Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



PAGE 2-B



REFERENCE COUNT:

6

THERE ARE 6 CITED REFERENCES AVAILABLE FOR THIS
RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L19 ANSWER 3 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1998:161806 HCAPLUS

DOCUMENT NUMBER: 128:291691

TITLE: Dentin phosphoprotein sequence motifs and molecular
modeling: conformational adaptations to mineral

crystals
AUTHOR(S): Dahlin, Stefan; Angstrom, Jonas; Linde, Anders
CORPORATE SOURCE: Dep. Oral Biochem., Goteborg Univ., Goteborg, Swed.
SOURCE: European Journal of Oral Sciences (1998), 106(Suppl.
1), 239-248
CODEN: EJOSFY; ISSN: 0909-8836
PUBLISHER: Munksgaard International Publishers Ltd.
DOCUMENT TYPE: Journal
LANGUAGE: English

AB Mol. modeling has been used to investigate structural features of oligopeptides derived from possible primary structure motifs in highly phosphorylated dentin phosphoprotein (PP-H), the predominant non-collagenous protein in dentin. It contains a large no. of aspartate (Asp) and phosphoserine (Pse) residues, the latter proposedly crucial for the PP-H function as a mineral nucleator. In this work, computer fitting and subsequent structural adaptation of model peptides, built exclusively from Asp and Pse, to the known crystal structures of hydroxyapatite (HAP) and octacalcium phosphate (OCP) were performed. The results show that, when considering conformational energies of fitted single strand oligo-peptides, either crystal will serve. Within a narrow range, fitting to OCP was slightly better fit to HAP. Energy differences between crystal-adapted and non-adapted freely minimized peptides showed that oligo(Pse-Asp) docked to either HAP or OCP were the energetically most favored adaptations. Fitting of minimized triple anti-parallel .beta.-strands of oligo(Pse-Asp) or oligo (Pse-Pse-Asp), motifs found in published sequences of rat, mouse, and bovine PP-H, revealed that a (001) crystal face of HAP, but most likely not OCP, may be formed by these .beta.-sheet models. The former motif is more advantageous in this respect.

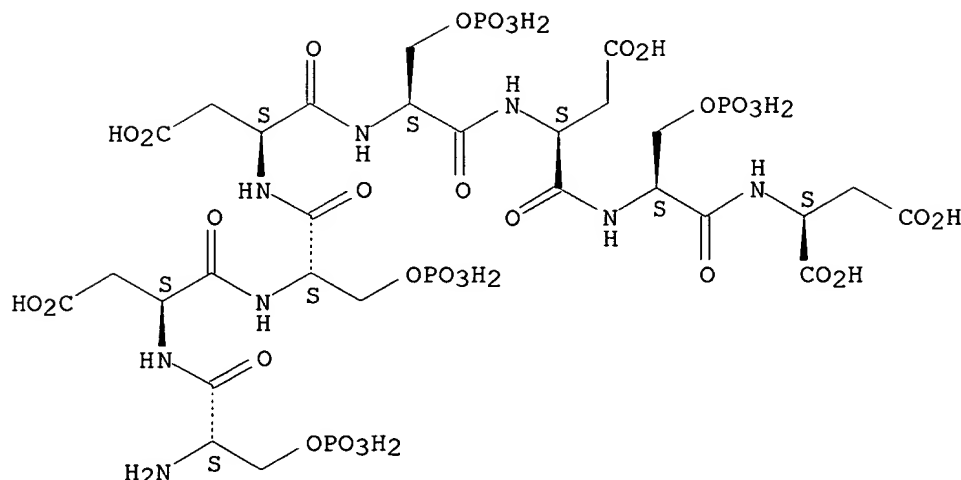
IT 206048-90-0

RL: BPR (Biological process); BSU (Biological study, unclassified); PRP (Properties); BIOL (Biological study); PROC (Process)
(dentin phosphoprotein sequence motifs and mol. modeling:
conformational adaptations to mineral crystals)

RN 206048-90-0 HCAPLUS

CN L-Aspartic acid, O-phosphono-L-seryl-L-.alpha.-aspartyl-O-phosphono-L-seryl-L-.alpha.-aspartyl-O-phosphono-L-seryl- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



L19 ANSWER 4 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1998:55543 HCAPLUS

DOCUMENT NUMBER: 128:110877

TITLE: Synthetic phosphopeptides for treating bone diseases

INVENTOR(S): Kumagai, Yoshinari; Otaka, Akira

PATENT ASSIGNEE(S): Big Bear Bio, Inc., USA

SOURCE: PCT Int. Appl., 45 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 2

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9800156	A1	19980108	WO 1997-US11426	19970630
W: AM, AU, BA, BG, BR, CA, CN, CZ, EE, FI, GE, HU, IL, IS, JP, KG, KR, LK, LT, LV, MD, MK, MN, MX, NO, NZ, PL, SG, SI, SK, TR, UA, UZ, VN, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
US 5837674	A	19981117	US 1996-675031	19960703
CA 2258661	AA	19980108	CA 1997-2258661	19970630
AU 9735871	A1	19980121	AU 1997-35871	19970630
AU 727675	B2	20001221		
EP 938326	A1	19990901	EP 1997-932409	19970630
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, NL, SE, PT, IE, FI				
JP 2001503452	T2	20010313	JP 1998-504399	19970630
PRIORITY APPLN. INFO.:				
			US 1996-675031	A 19960703
			WO 1997-US11426	W 19970630

AB Phosphopeptides which significantly reduce bone loss or weakening are provided. A method for treating or preventing any conditions assocd. with bone loss or weakening by administering the phosphopeptides by oral or injectable means is also provided. After age 35, bone mass, mineral content and mech. strength of the bone begin declining gradually. The relationship between bone mass and age is shown. Examples of prevention of bone loss in an osteoporosis model are given for peptides such as Pse-Gly-Pse-Gly-Pse-Gly (Pse = O-phosphoserine).

IT 201660-41-5 201660-42-6 201660-43-7

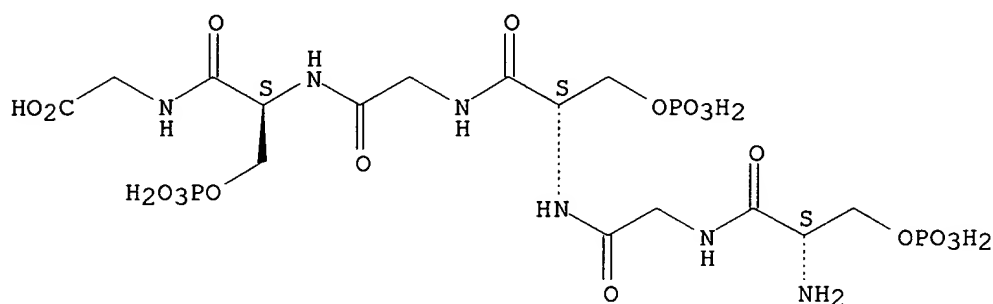
RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); THU (Therapeutic use); BIOL (Biological study); USES (Uses) .

(synthetic phosphopeptides for treating bone diseases)

RN 201660-41-5 HCAPLUS

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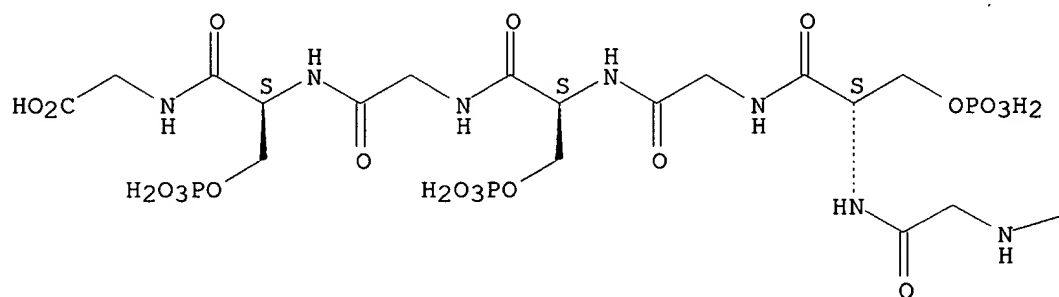
Absolute stereochemistry.



RN 201660-42-6 HCAPLUS

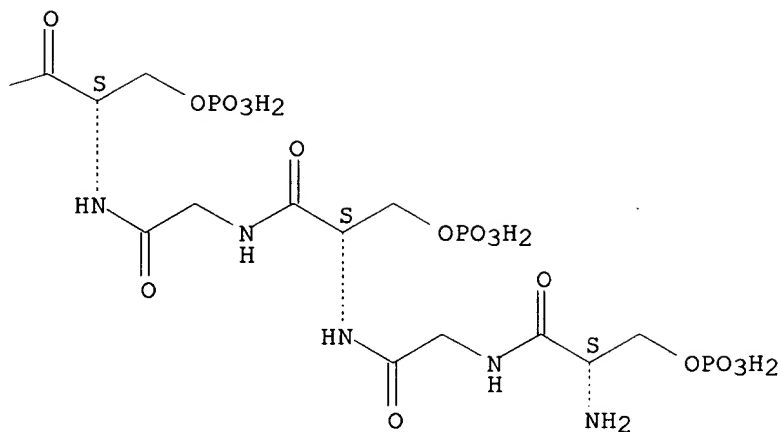
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Absolute stereochemistry.



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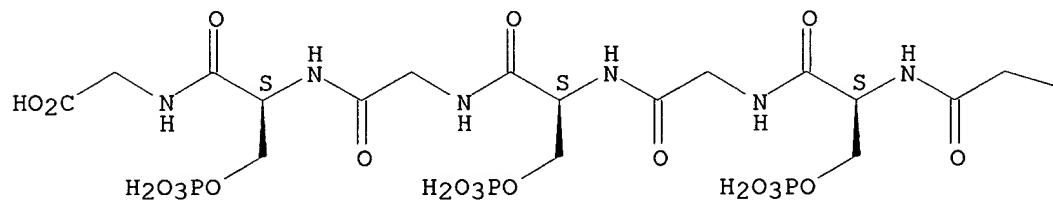


RN 201660-43-7 HCAPLUS

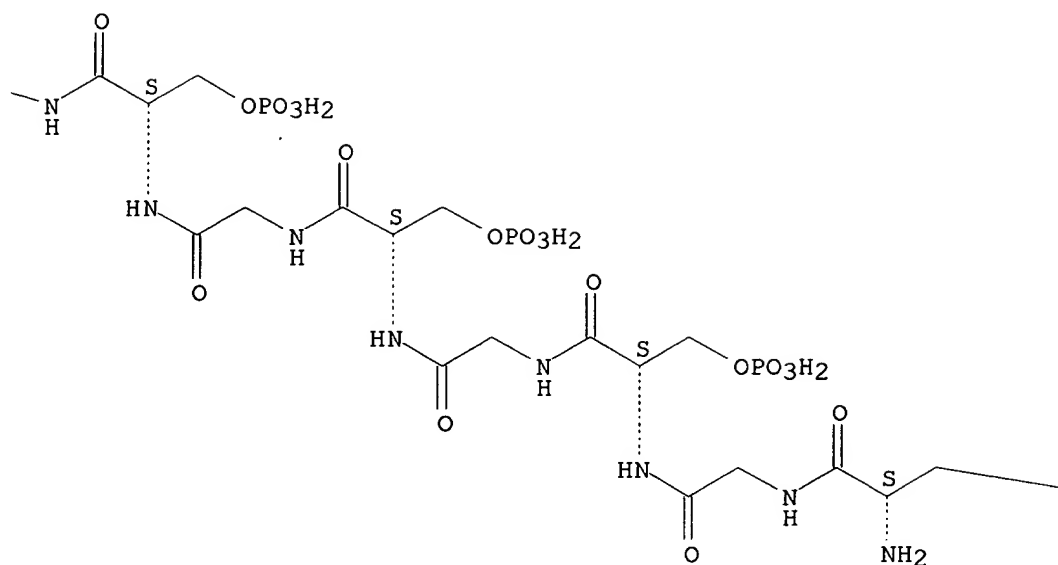
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Absolute stereochemistry.

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PAGE 1-C

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L19 ANSWER 5 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1997:457199 HCAPLUS

DOCUMENT NUMBER: 127:172781

TITLE: High-affinity binding of the Drosophila Numb
phosphotyrosine-binding domain to peptides containing
a Gly-Pro-(p)Tyr motifAUTHOR(S): Li, Shun-Cheng; Songyang, Zhou; Vincent, Sebastien J.
F.; Zwahlen, Catherine; Wiley, Sandra; Cantley, Lewis;
Kay, Lewis E.; Forman-Kay, Julie; Pawson, Tony

CORPORATE SOURCE: Program Molecular Biology Cancer, Samuel Lunenfeld
Research Institute, Mount Sinai Hospital, Toronto, ON,
M5G 1X5, Can.

SOURCE: Proceedings of the National Academy of Sciences of the
United States of America (1997), 94(14), 7204-7209
CODEN: PNASA6; ISSN: 0027-8424

PUBLISHER: National Academy of Sciences

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The phosphotyrosine-binding (PTB) domain is a recently identified protein
module that has been characterized as binding to phosphopeptides contg. an
NPXpY motif (X = any amino acid). We describe here a novel peptide
sequence recognized by the PTB domain from Drosophila Numb (dNumb), a
protein involved in cell fate detn. and asym. cell division during the
development of the Drosophila nervous system. Using a Tyr-oriented
peptide library to screen for ligands, the dNumb PTB domain was found to
bind selectively to peptides contg. a YIGPY.phi. motif (.phi. represents a
hydrophobic residue). A synthetic peptide contg. this sequence bound
specifically to the isolated dNumb PTB domain in soln. with a dissocn.
const. (Kd) of $5.78 \pm 0.74 \mu\text{M}$. Interestingly, the affinity of this
peptide for the dNumb PTB domain was increased ($K_d = 1.41 \pm 0.10 \mu\text{M}$)
when the second tyrosine in the sequence was phosphorylated. Amino acid
substitution studies of the phosphopeptide demonstrated that a core motif
of sequence GP(p)Y is required for high-affinity binding to the dNumb PTB
domain. NMR expts. performed on isotopically labeled protein complexed
with either Tyr- or pTyr-contg. peptides suggest that the same set of
amino acids in the dNumb PTB domain is involved in binding both
phosphorylated and nonphosphorylated forms of the peptide. The in vitro
selectivity of the dNumb PTB domain is therefore markedly different from
those of the Shc and IRS-1 PTB domains, in that it interacts
preferentially with a GP(p)Y motif, rather than NPXpY, and does not
absolutely require ligand phosphorylation for binding. Our results
suggest that the PTB domain is a versatile protein module, capable of
exhibiting varied binding specificities.

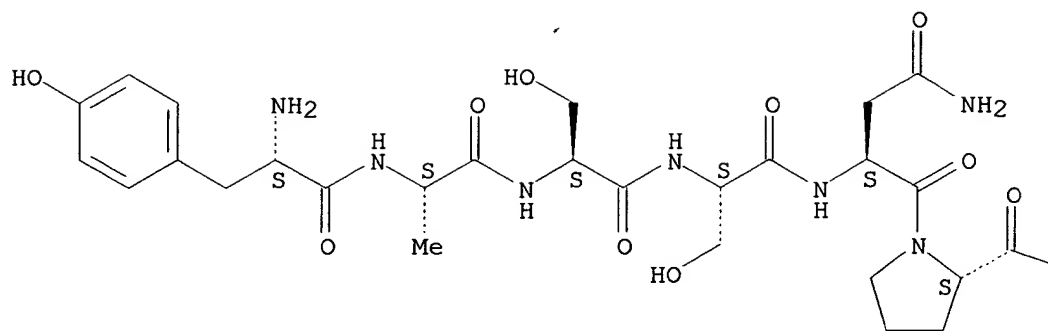
IT **186312-67-4**
RL: BPR (Biological process); BSU (Biological study, unclassified); PRP
(Properties); BIOL (Biological study); PROC (Process)
(high-affinity binding of the Drosophila Numb phosphotyrosine-binding
domain to peptides contg. a Gly-Pro-(p)Tyr motif)

RN 186312-67-4 HCAPLUS

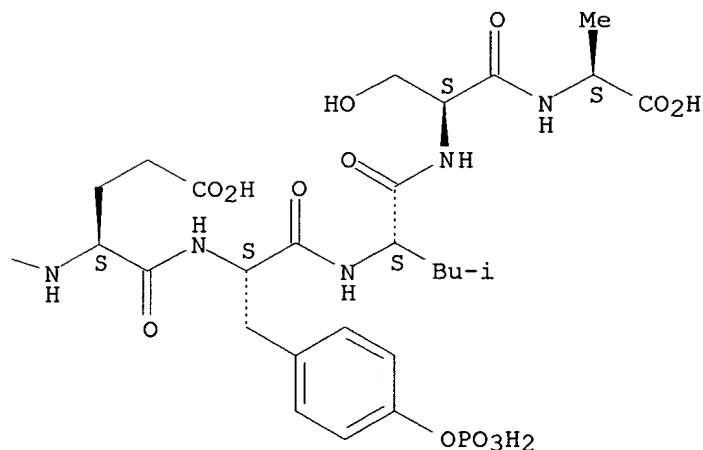
CN L-Alanine, L-tyrosyl-L-alanyl-L-seryl-L-seryl-L-asparaginyl-L-prolyl-L-
.alpha.-glutamyl-O-phosphono-L-tyrosyl-L-leucyl-L-seryl- (9CI) (CA INDEX
NAME)

Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



L19 ANSWER 6 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1997:377883 HCAPLUS

DOCUMENT NUMBER: 126:338844

TITLE: Phosphotyrosine-containing peptide inhibitors of a phosphotyrosine-binding domain containing protein for the treatment of Shc-dependent conditions

INVENTOR(S): Van Der Geer, Peter; Wiley, Sandra; Gish, Gerald; Pawson, Tony; Toma, Kazunori

PATENT ASSIGNEE(S): Mount Sinai Hospital Corporation, Can.; Asahi Chemical Industry Co., Ltd.; Van Der Geer, Peter; Wiley, Sandra; Gish, Gerald; Pawson, Tony; Toma, Kazunori

SOURCE: PCT Int. Appl., 71 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
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WO 9715318	A1	19970501	WO 1996-US17080	19961024
W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, US, US, UZ, VN, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
RW: KE, LS, MW, SD, SZ, UG, AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ				
CA 2235756	AA	19970501	CA 1996-2235756	19961024
AU 9675211	A1	19970515	AU 1996-75211	19961024
EP 874639	A1	19981104	EP 1996-937741	19961024
R: CH, DE, FR, GB, LI				
JP 11514381	T2	19991207	JP 1996-516793	19961024
US 6028053	A	20000222	US 1998-51934	19980422
PRIORITY APPLN. INFO.:			US 1995-5944P	P 19951027
			US 1996-10384P	P 19960122
			US 1996-11799P	P 19960220
			WO 1996-US17080	W 19961024

AB Phosphotyrosine-contg. peptides derived from the Asn-Pro-X-phosphoTyr motif found in phosphotyrosine-contg. proteins are described. These peptides interfere with the binding of proteins to the phosphotyrosine-binding domains and can be used to control signal transduction processes, esp. those involving SH domain-contg. proteins. The peptides may be prepd. by std. chem. or biol. by expression of the corresponding gene or incorporation into a fusion protein.

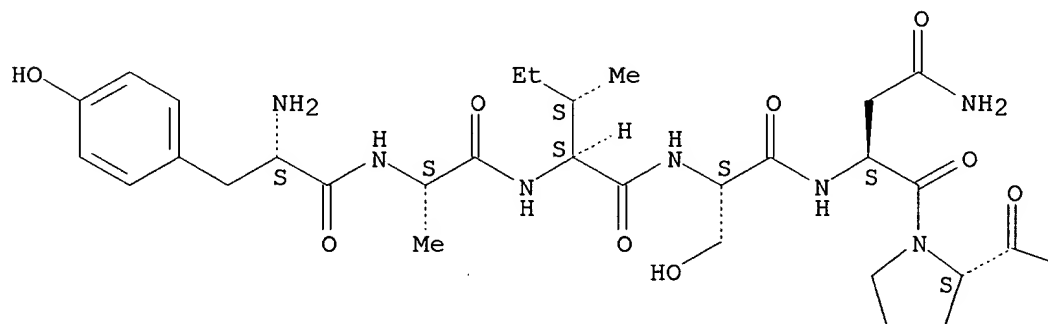
IT **186312-68-5**
 RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); PRP (Properties); THU (Therapeutic use); BIOL (Biological study); USES (Uses)
 (amino acid sequence, inhibition of Shc binding to growth factor receptors and; phosphotyrosine-contg. peptide inhibitors of phosphotyrosine-binding domain contg. protein for treatment of Shc-dependent conditions)

RN 186312-68-5 HCAPLUS

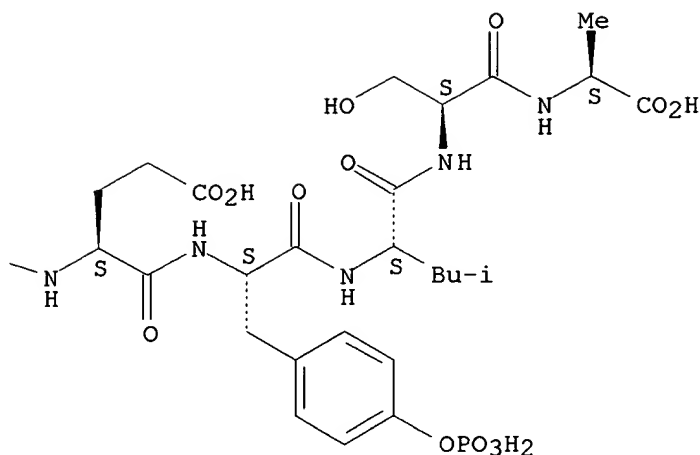
CN L-Alanine, L-tyrosyl-L-alanyl-L-isoleucyl-L-seryl-L-asparaginyl-L-prolyl-L-.alpha.-glutamyl-O-phosphono-L-tyrosyl-L-leucyl-L-seryl- (9CI) (CA INDEX NAME)

Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



L19 ANSWER 7 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1997:21554 HCAPLUS

DOCUMENT NUMBER: 126:114763

TITLE: Characterization of the phosphotyrosine-binding domain of the Drosophila Shc protein

AUTHOR(S): Li, Shun-Cheng; Lai, Ka-Man Venus; Gish, Gerald D.; Parris, Wendy E.; van der Geer, Peter; Forman-Kay, Julie; Pawson, Tony

CORPORATE SOURCE: Samuel Lunenfeld Res. Inst., Mt. Sinai Hosp., Toronto, ON, M5G 1X5, Can.

SOURCE: Journal of Biological Chemistry (1996), 271(50), 31855-31862

CODEN: JBCHA3; ISSN: 0021-9258

PUBLISHER: American Society for Biochemistry and Molecular Biology

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The phosphotyrosine-binding (PTB) domain of Drosophila Shc (dShc) binds in vitro to phosphopeptides contg. the sequence motif NPXpY, and phys. assoc. with the activated Drosophila epidermal growth factor receptor homolog (DER) in vivo. The structural elements, specificity and binding kinetics of the dShc PTB domain have now been characterized. The dShc PTB domain appeared similar to the insulin-like receptor substrate-1 PTB domain in secondary structure as suggested by Fourier transform IR spectroscopy. Surface plasmon resonance measurements indicated that the dShc PTB domain bound with high affinity to phosphopeptides (Der) derived from the Tyr1228 site of the DER receptor. The kinetics of the dShc PTB domain-Der phosphopeptide interaction differed from those of a typical SH2 domain-ligand interaction, in that the PTB domain displayed slower on/off rates. Competition binding assays using truncated versions of the Der peptides revealed that high affinity binding to the dShc PTB domain requires, in addn. to the NPXpY motif, the presence of hydrophobic residues at both positions -5 and -7 relative to phosphotyrosine. The dShc PTB domain showed a similar binding specificity to the human Shc (hShc) PTB domain, but subtle differences were noted; such that the hShc

PTB domain bound preferentially to a phosphopeptide from the mammalian nerve growth factor receptor, whereas the dShc PTB domain bound preferentially to phosphopeptides from the Drosophila DER receptor. The invertebrate dShc PTB domain therefore possesses a binding specificity for tyrosine-phosphorylated peptides that is optimally suited for recognition of the activated DER receptor.

IT 186312-67-4 186312-68-5

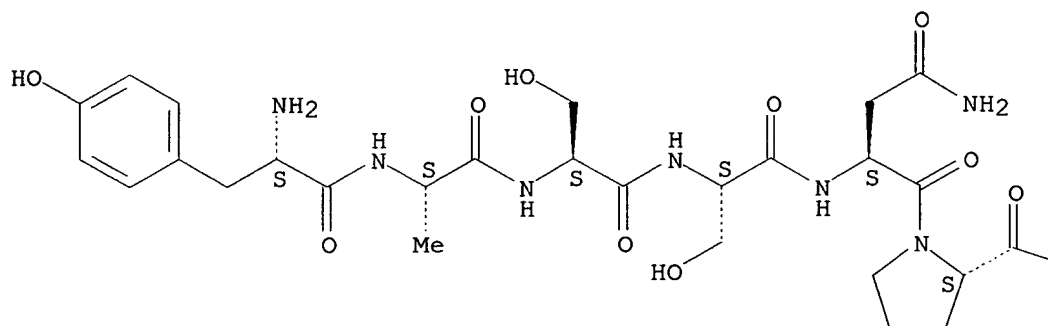
RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process)
(characterization of phosphotyrosine-binding domain of Shc protein)

RN 186312-67-4 HCAPLUS

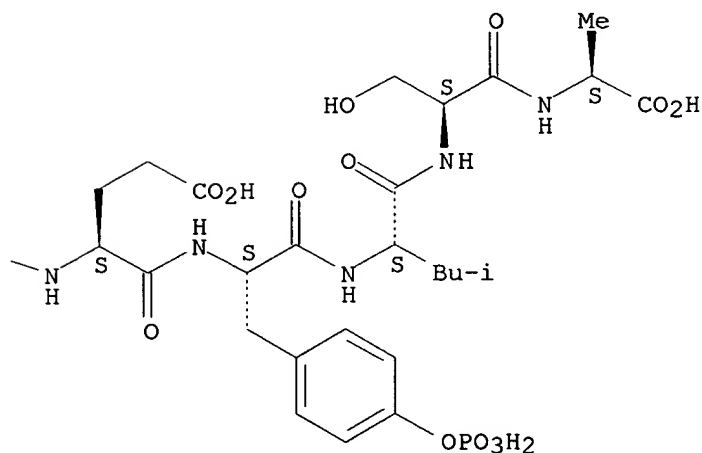
CN L-Alanine, L-tyrosyl-L-alanyl-L-seryl-L-seryl-L-asparaginyl-L-prolyl-L-.alpha.-glutamyl-O-phosphono-L-tyrosyl-L-leucyl-L-seryl- (9CI) (CA INDEX NAME)

Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



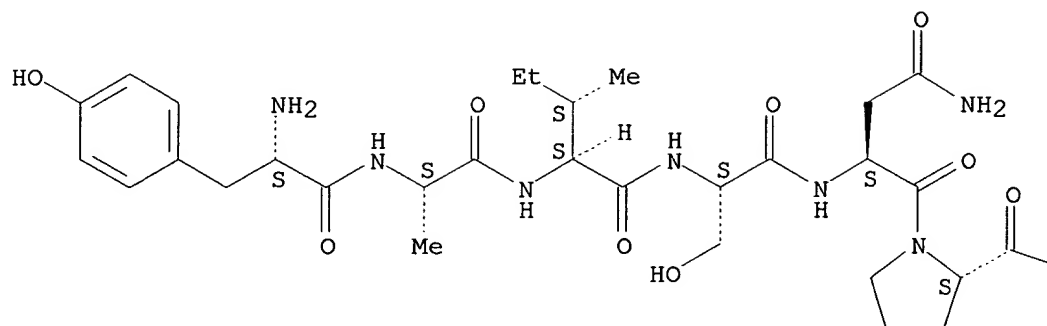
RN 186312-68-5 HCAPLUS

CN L-Alanine, L-tyrosyl-L-alanyl-L-isoleucyl-L-seryl-L-asparaginyl-L-prolyl-L-.alpha.-glutamyl-O-phosphono-L-tyrosyl-L-leucyl-L-seryl- (9CI) (CA INDEX NAME)

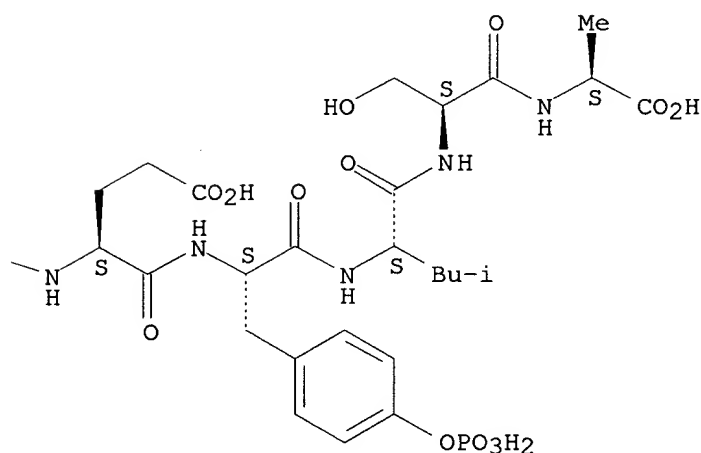
NAME)

Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



L19 ANSWER 8 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1996:153236 HCAPLUS

DOCUMENT NUMBER: 124:220717

TITLE: Systematic mapping of potential binding sites for Shc and Grb2 SH2 domains on insulin receptor substrate-1 and the receptors for insulin, epidermal growth factor, platelet-derived growth factor, and fibroblast growth factor

AUTHOR(S): Ward, Colin W.; Gough, Keith H.; Rashke, Melisa; Wan, Soo San; Tribbick, Gordon; Wang, Jian-xin

CORPORATE SOURCE: Division Biomolecular Engineering, CSIRO, Parkville, 3052, Australia

SOURCE: Journal of Biological Chemistry (1996), 271(10), 5603-9

CODEN: JBCHA3; ISSN: 0021-9258

PUBLISHER: American Society for Biochemistry and Molecular
Biology
DOCUMENT TYPE: Journal
LANGUAGE: English

AB Multipin peptide synthesis has been employed to produce biotinylated 11-mer phosphopeptides that account for every tyrosine residue in insulin receptor substrate-1 (IRS-1) and the cytoplasmic domains of the insulin-, epidermal growth factor-, platelet-derived growth factor- and basic fibroblast growth factor receptors. These phosphopeptides have been screened for their capacity to bind to the SH2 domains of Shc and Grb in a soln. phase ELISA. The data revealed new potential Grb2 binding sites at Tyr-1114 (epidermal growth factor receptor (EGFR) C-tail); Tyr-743 (platelet-derived growth factor receptor (PDGFR) insert region), Tyr-1110 from the E-helix of the catalytic domain of insulin receptor (IR), and Tyr-47, Tyr-939, and Tyr-727 in IRS-1. None of the phosphopeptides from the juxtamembrane or C-tail regions of IR bound Grb2 significantly, and only one phosphopeptide from the basic fibroblast growth factor receptor (Tyr-556) bound Grb2 but with medium strength. Tyr-1068 and -1086 from the C-tail of EGFR, Tyr-684 from the kinase insert region of PDGFR, and Tyr-895 from IRS-1 were confirmed as major binding sites for the Grb2 SH2 domain. With regard to Shc binding, the data revealed new potential binding sites at Tyr-703 and Tyr-789 from the catalytic domain of EGFR and at Tyr-557 in the juxtamembrane region of PDGFR. It also identified new potential Shc binding sites at Tyr-764, in the C-tail of basic fibroblast growth factor receptor, and Tyr-960, in the juxtamembrane of IR, a residue previously known to be required for Shc phosphorylation in response to insulin. The study confirmed the previous identification of Tyr-992 and Tyr-1173 in the C-tail of EGFR and several phosphopeptides from the PDGFR as medium strength binding sites for the SH2 domain of Shc. None of the 34 phosphopeptides from IRS-1 bound Shc strongly, although Tyr-690 showed medium strength binding. The specificity characteristics of the SH2 domains of Grb2 and Shc are discussed. This systematic peptide mapping strategy provides a way of rapidly scanning candidate proteins for potential SH2 binding sites as a first step to establishing their involvement in kinase-mediated signaling pathways.

IT 174660-15-2

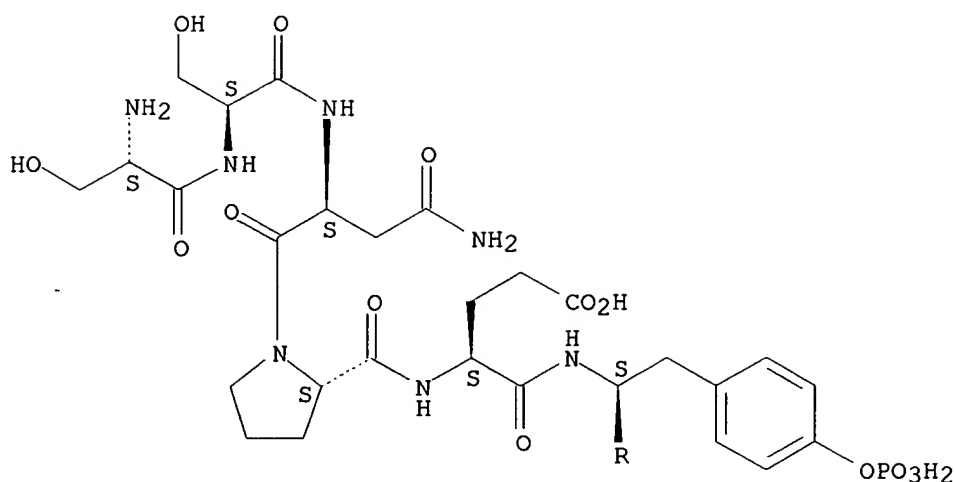
RL: BPR (Biological process); BSU (Biological study, unclassified); PRP (Properties); BIOL (Biological study); PROC (Process)
(Shc and Grb2 SH2 domain binding site mapping on insulin receptor substrate-1 and insulin receptor and growth factor receptors)

RN 174660-15-2 HCAPLUS

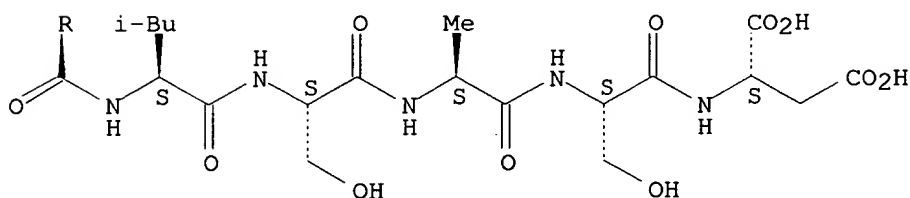
CN L-Aspartic acid, N-[N-[N-[N-[N-[O-phosphono-N-[N-[1-[N2-(N-L-seryl-L-seryl)-L-asparaginy]-L-prolyl]-L-.alpha.-glutamyl]-L-tyrosyl]-L-leucyl]-L-seryl]-L-alanyl]-L-seryl]- (9CI) (CA INDEX NAME)

Absolute stereochemistry.

PAGE 1-A



PAGE 2-A

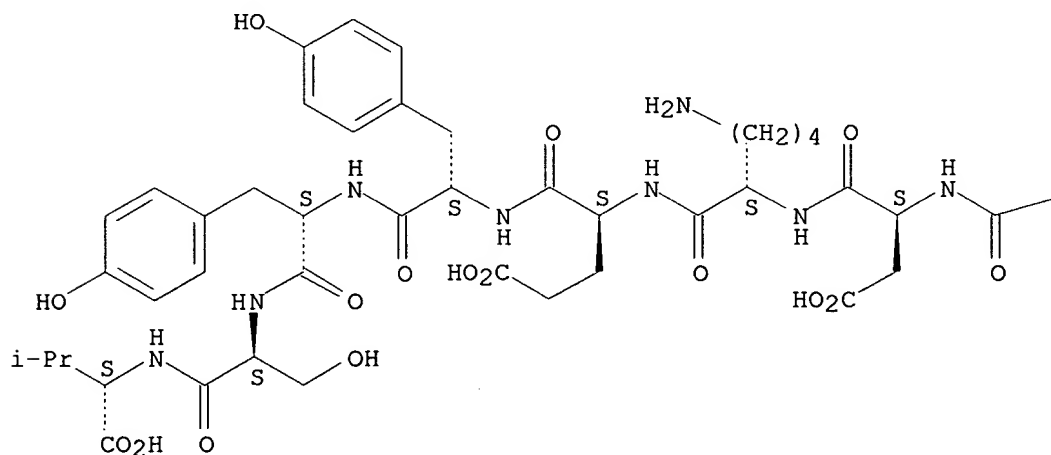


L19 ANSWER 9 OF 24 HCAPLUS COPYRIGHT 2002 ACS
 ACCESSION NUMBER: 1995:541384 HCAPLUS
 DOCUMENT NUMBER: 122:282231
 TITLE: Peptide inhibitors of mitogenesis and motogenesis
 INVENTOR(S): Comoglio, Paolo; Ponzetto, Carola
 PATENT ASSIGNEE(S): Farmitalia Carlo Erba S.r.L., Italy
 SOURCE: PCT Int. Appl., 86 pp.
 CODEN: PIXXD2
 DOCUMENT TYPE: Patent
 LANGUAGE: English
 FAMILY ACC. NUM. COUNT: 1
 PATENT INFORMATION:

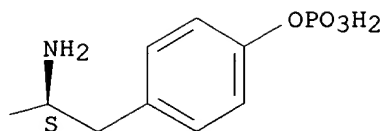
PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9501376	A1	19950112	WO 1994-EP1943	19940615
W: AU, BB, BG, BR, BY, CA, CN, CZ, FI, HU, JP, KP, KR, KZ, LK, LV, MG, MN, MW, NO, NZ, PL, RO, RU, SK, UA, UZ, VN				
RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
CA 2142713	AA	19950112	CA 1994-2142713	19940615
AU 9470718	A1	19950124	AU 1994-70718	19940615
AU 670704	B2	19960725		
EP 662090	A1	19950712	EP 1994-919632	19940615

Absolute stereochemistry.

PAGE 1-A



PAGE 1-B



L19 ANSWER 10 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1991:492927 HCAPLUS

DOCUMENT NUMBER: 115:92927

TITLE: Synthesis of casein-related peptides and phosphopeptides. XIV. Solid-phase synthesis of Glu-Ser(P)-Leu through the use of protected Boc-Ser(PO3R2)-OH derivatives

AUTHOR(S): Perich, John W.; Valerio, Robert M.; Alewood, Paul F.; Johns, R. B.

CORPORATE SOURCE: Sch. Chem., Univ. Melbourne, Parkville, 3052, Australia

SOURCE: Australian Journal of Chemistry (1991), 44(6), 771-8
CODEN: AJCHAS; ISSN: 0004-9425

DOCUMENT TYPE: Journal

LANGUAGE: English

AB A solid-phase method is described for the synthesis of O-phosphoseryl-contg. peptides H-Glu-Ser(PO3R2)-Leu-OH (I; R = H, Et) and H-Ser(PO3H2)-Leu-OH with Boc-Ser(PO3R2)-OH (Boc = Me3CO2C; R = Et, Ph, CMe3) and PhCMe2O2-Ser[PO3(CH2Ph)2]-OH for the incorporation of the phosphorylated seryl residue.

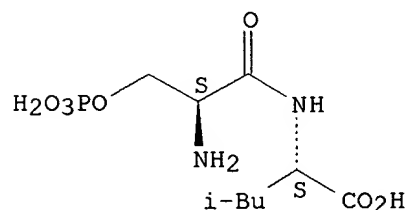
IT 135432-35-8P 135432-36-9P

RL: SPN (Synthetic preparation); PREP (Preparation)
(prepn. of)

RN 135432-35-8 HCAPLUS

CN L-Leucine, N-(O-phosphono-L-seryl)-, monohydrobromide (9CI) (CA INDEX NAME)

Absolute stereochemistry.



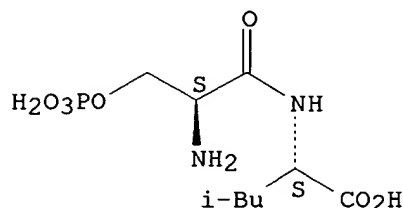
HBr

RN 135432-36-9 HCAPLUS
 CN L-Leucine, N-(O-phosphono-L-seryl)-, mono(trifluoroacetate) (9CI) (CA
 INDEX NAME)

CM 1

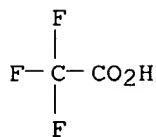
CRN 6665-27-6
 CMF C9 H19 N2 O7 P

Absolute stereochemistry.



CM 2

CRN 76-05-1
 CMF C2 H F3 O2



L19 ANSWER 11 OF 24 HCAPLUS COPYRIGHT 2002 ACS
 ACCESSION NUMBER: 1991:186053 HCAPLUS
 DOCUMENT NUMBER: 114:186053
 TITLE: Synthesis of casein-related peptides and
 phosphopeptides. VIII. The synthesis of
 Ser(P)-containing peptides by the use of
 Z-Ser(PO3R2)-OH derivatives
 AUTHOR(S): Perich, John W.; Alewood, Paul F.; Johns, R. B.
 CORPORATE SOURCE: Sch. Chem., Univ. Melbourne, Parkville, 3052,
 Australia
 SOURCE: Australian Journal of Chemistry (1991), 44(2), 253-63
 CODEN: AJCHAS; ISSN: 0004-9425
 DOCUMENT TYPE: Journal
 LANGUAGE: English
 OTHER SOURCE(S): CASREACT 114:186053
 AB The synthesis of five Z-Ser(PO3R2)-OH (I; Z = PhCH2O2C; R = Ph, Et, Me,
 CH2Ph, CMe3) is described by a simple three-step synthetic procedure which
 features the use of either di-Ph or dialkyl phosphorochloridate/pyridine
 or dialkyl N,N-diethylphosphoramidite/1H-tetrazole-m-chloroperoxybenzoic
 acid for the phosphorylation of the serine hydroxy group. The benzyl
 phosphate deriv. I (R = CH2Ph) was used in the benzyloxycarbonyl mode of
 peptide synthesis for the prepn. of Z-Ser[PO3(CH2Ph)2]-X-OCH2Ph [X = Leu,

Ser[PO₃(CH₂Ph)₂] which were deprotected by palladium-catalyzed hydrogenolysis to give the corresponding phosphoserine dipeptides in high yields and high purity.

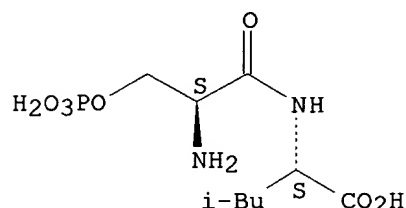
IT **6665-27-6P**

RL: SPN (Synthetic preparation); PREP (Preparation)
(prepn. of)

RN 6665-27-6 HCAPLUS

CN L-Leucine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



L19 ANSWER 12 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1991:186052 HCAPLUS

DOCUMENT NUMBER: 114:186052

TITLE: Synthesis of casein-related peptides and phosphopeptides. VII. The efficient synthesis of Ser(P)-Containing peptides by the use of Boc-Ser(PO₃R₂)-OH derivatives

AUTHOR(S): Perich, John W.; Alewood, Paul F.; Johns, R. B.

CORPORATE SOURCE: Sch. Chem., Univ. Melbourne, Parkville, 3052, Australia

SOURCE: Australian Journal of Chemistry (1991), 44(2), 233-52
CODEN: AJCHAS; ISSN: 0004-9425

DOCUMENT TYPE: Journal

LANGUAGE: English

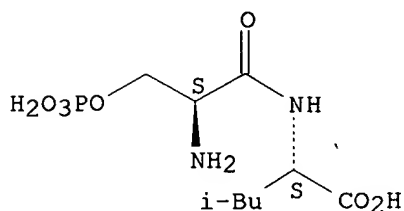
OTHER SOURCE(S): CASREACT 114:186052

AB A new approach to the synthesis of Ser(PO₃R₂)-contg. peptides by using five protected Boc-Ser(PO₃R₂)-OH (I; Boc = Me₃CO₂C; R = Ph, Et, Me, CH₂Ph, CMe₃) derivs. is described. I are prepd. by a simple three-step synthetic procedure which features the use of either di-Ph or dialkyl phosphorochloridate/pyridine, or dialkyl N,N-diethylphosphoramidite/1H-tetrazole-m-chloroperoxybenzoic acid for the phosphorylation of the serine hydroxy group. I were utilized in the Boc mode of peptide synthesis with the use of the mixed anhydride coupling procedure, and led to the synthesis of the protected tripeptides, Boc-Glu(OCH₂Ph)-Ser(PO₃R₂)-Leu-OCH₂Ph in high yield and purity. In peptide deprotection studies, the Ph and benzyl phosphate protecting groups were readily removed from the protected tripeptides by hydrogenolysis and gave the phosphoserine tripeptide in near quant. yield. The tert-Bu phosphate groups were cleaved from Boc-Ser[PO₃(CMe₃)₂]-Leu-OCH₂Ph by mild acidolysis and gave the phosphoserine dipeptide in quant. yield. However, attempts to cleave the Et or Me phosphate groups by acidolytic or silylitic treatment of the corresponding tripeptides resulted in decompn. of the O-phosphoseryl residue.

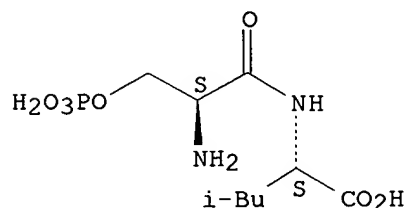
IT **6665-27-6P**

RL: SPN (Synthetic preparation); PREP (Preparation)
(prepn. of)

Absolute stereochemistry.



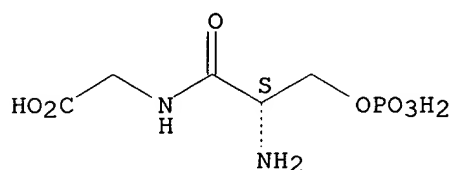
Absolute stereochemistry.



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L19  ANSWER 14 OF 24  HCAPLUS  COPYRIGHT 2002 ACS
ACCESSION NUMBER:      1987:85018  HCAPLUS
DOCUMENT NUMBER:       106:85018
TITLE:                 Solid-phase synthesis of phosphopeptides:  synthesis
                        of phosphopeptides from the carboxyl-terminus of
                        rhodopsin
```


AUTHOR(S): Arendt, Anatol; Hargrave, Paul A.
 CORPORATE SOURCE: Dep. Ophthalmol., Univ. Florida, Gainesville, FL,
 32610, USA
 SOURCE: Pept.: Struct. Funct., Proc. Am. Pept. Symp., 9th
 (1985), 237-40
 CODEN: 54ZNAJ
 DOCUMENT TYPE: Conference
 LANGUAGE: English
 AB Me3CO2CNHCH(CO2H)CHOP(O)(OPh)2 (R = H, Me) were prepd. and used in the
 solid-phase synthesis of phosphorylated serine (PSer) and threonine (PThr)
 contg. peptides H-PSer-Gly-OH, H-Val-PSer-Lys-OH, H-Glu-PThr-PSer-Gln-Val-
 OH, and H-Val-Ser-Lys-Thr-Glu-PThr-PSer-Gln-Val-OH.
 IT **6665-42-5P**
 RL: SPN (Synthetic preparation); PREP (Preparation)
 (prepn. of)
 RN 6665-42-5 HCAPLUS
 CN Glycine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



L19 ANSWER 15 OF 24 HCAPLUS COPYRIGHT 2002 ACS
 ACCESSION NUMBER: 1984:2581 HCAPLUS
 DOCUMENT NUMBER: 100:2581
 TITLE: Tripeptidyl aminopeptidase in the extralysosomal
 fraction of rat liver
 AUTHOR(S): Baaloe, Ros Mari; Ragnarsson, Ulf; Zetterqvist,
 Oerjan
 CORPORATE SOURCE: Biomed. Cent., Univ. Uppsala, Uppsala, S-751 23, Swed.
 SOURCE: Journal of Biological Chemistry (1983), 258(19),
 11622-8
 CODEN: JBCHA3; ISSN: 0021-9258
 DOCUMENT TYPE: Journal
 LANGUAGE: English
 AB Tripeptidyl aminopeptidase (I), an enzyme which removes tripeptides from
 the free, N-terminal end of oligopeptides, was detected in the
 extralysosomal fraction of rat liver. I was partially purified by
 Sepharose CL-4B and DEAE-cellulose chromatog. The pH optimum was in the
 neutral range and the apparent native mol. wt. was >106, as judged by
 Sepharose chromatog. I cleaved the phosphopeptide, Gly-Val-Leu-Arg-Arg-
 Ala-Ser(P)-Val-Ala, 1st at the Leu-Arg bond and then at the Ala-Ser(P)
 bond. The cleavage of the former bond was inhibited by
 Arg-Arg-Ala-Ser(P)-Val-Ala (II), which indicated that both bonds were
 cleaved by the same enzyme. The Km for II was 0.01 mM at pH 6.5-7.5.
 Val-Leu-Arg-Arg-Ala-Ser(P)-Val-Ala (III) and Leu-Arg-Arg-Ala-Ser(P)-Val-
 Ala were poor substrates. III was, however, an efficient inhibitor. The
 Ala-Ser(P) bond of Arg-Arg-Ala-Ser(P)-Val (IV) was cleaved at the same
 rate as that of II. I was active also with the unphosphorylated peptides
 corresponding to II and IV and tolerated the substitution of lysine for

the N-terminal arginine of the latter peptide. Substitution of guanidovaleric acid for the N-terminal arginine of IV and of guanidovaleric acid or .epsilon.-aminohexanoic acid for the N-terminal arginine of unphosphorylated IV reduced the rate of hydrolysis to insignificant levels, demonstrating the importance of a free N-terminus. The results thus provide evidence of a unique I.

IT **88169-78-2**

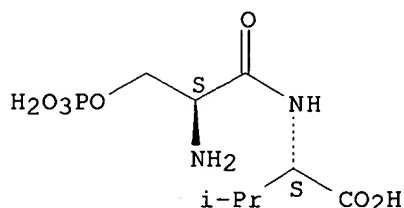
RL: FORM (Formation, nonpreparative)

(formation of, by tripeptidyl aminopeptidase of liver)

RN 88169-78-2 HCAPLUS

CN L-Valine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



L19 ANSWER 16 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1971:482043 HCAPLUS

DOCUMENT NUMBER: 75:82043

TITLE: Ultraviolet, visible, and infrared spectroscopic studies of the interaction of hydroxocobalamin with .alpha.-amino acids and peptides

AUTHOR(S): Heathcote, J. G.; Moxon, G. H.; Slifkin, M. A.

CORPORATE SOURCE: Univ. Salford, Salford, Engl.

SOURCE: Spectrochim. Acta, Part A (1971), 27(8), 1391-408
CODEN: SAMCAS

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Evidence is given for the formation of mol. complexes between vitamin B12b and certain .alpha.-amino acids, with a mol. ratio of 1.1. Infrared data show that charge donation occurs from the N of the amino acid, in the unionized form. Rate consts. for complex formation are also given. Evidence is presented for the formation of mol. complexes between hydroxocobalamin (vitamin B12b) and dipeptides, polypeptides, or proteins. It is obsd. that hydroxocobalamin forms a complex with glycine more readily than with other .alpha.-amino acids. This extends to peptides also; thus, peptides contg. N-terminal glycine complex more strongly than those contg. other N-terminal amino acids. Nevertheless, other amino acids, in particular the 2nd amino acid in the chain, play a part in detg. complexing ability. There is a significant decrease in assocn. as one goes down the series Ala-Gly, Ala-Ala, Ala-Ser. Steric effects also appear to be important.

IT **33897-83-5**

RL: PRP (Properties)

(spectrum of)

RN 33897-83-5 HCAPLUS

CN Cobinamide, dihydroxide, dihydrogen phosphate (ester), mono(inner salt), 3'-ester with 5,6-dimethyl-1-.alpha.-D-ribofuranosylbenzimidazole, compd. with N-L-serylglycine (1:1) (8CI) (CA INDEX NAME)

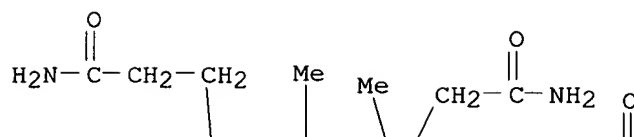
CM 1

CRN 13422-51-0

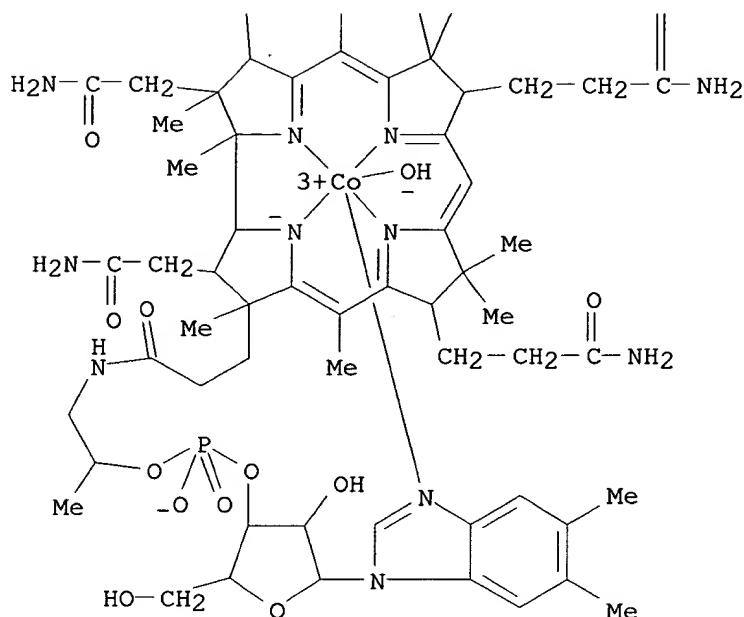
CMF C62 H89 Co N13 O15 P

CCI CCS

PAGE 1-A



PAGE 2-A

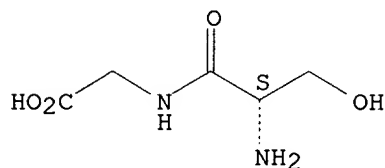


CM 2

CRN 687-63-8

CMF C5 H10 N2 O4

Absolute stereochemistry.



L19 ANSWER 17 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1966:412631 HCAPLUS

DOCUMENT NUMBER: 65:12631

ORIGINAL REFERENCE NO.: 65:2346f-h,2347a-h,2348a-h,2349a-e

TITLE: Synthesis of phosphopeptides. V. Further dipeptides, tripeptides, and O-phosphorylated derivatives of L-serine

AUTHOR(S): Folsch, Georg

CORPORATE SOURCE: Univ. Goteborg, Swed.

SOURCE: Acta Chem. Scand. (1966), 20(2), 459-73

DOCUMENT TYPE: Journal

LANGUAGE: English

AB cf. CA 57, 4794h. Various phosphorylated di- and tripeptides as well as the corresponding phosphate-free peptides were synthesized via N-benzyloxycarbonyl peptide benzyl esters. Protected di- and tripeptide derivs. of .beta.-Bzl-Asp-Set (Bzl = benzyl) all underwent a rapid

cyclization to aspartimides under weakly alk. conditions, e.g. when dissolved in C₅H₅N-H₂O, whereas the corresponding derivs. of .gamma.-Bzl-Glu-Ser were stable under the same conditions. Some further new derivs. of serine and threonine were prepd. and studied. (All amino acids used have the L-configuration, except glycine. Z = PhCH₂O₂C; SerP = phosphorylserine = H₂NCH(CH₂OPO₃H₂)-CO₂H; DCCI = dicyclohexylcarbodiimide; THF = tetrahydrofuran). Z-Aspartic acid .beta.-Bzl ester (Ia) (Benoiton, CA 57, 9943g) (3.6 g.), 3.6 g. serine Bzl ester benzenesulfonate (I) (CA 56, 7418e), 1.4 ml. Et₃N, and 2.1 g. DCCI in 100 ml. 1:1 THF-MeCN shaken 4 hrs. at 4.degree. and 18 hrs. at room temp. and the soln. filtered, concd. in vacuo at 40.degree., and poured into 1 l. ice H₂O with stirring gave 4.4 g. Z-.alpha.-Asp-Ser-(Bzl)₂ (II), m. 129.degree. (EtOAc-Et₂O), [.alpha.]_{26D} 6.9.degree. (c 5.8, AcOH), [.alpha.]_{26D} 0.degree. (c 3.0, C₅H₅N). The optical rotation of II (300 mg.) increased steadily when dissolved in 10.5 ml. C₅H₅N and 4.5 ml. H₂O. After 5 days at 26.degree., the soln. was evapd. in vacuo and the residue triturated with Et₂O to give 120 mg. solid, m. 118-22.degree., which was hydrogenolyzed in tert-BuOH-H₂O. Paper chromatographic analysis of the product formed showed the presence of 3 ninhydrin-pos. compds. One of the spots gave the normal purple color and had the R_f 0.50 [2:1:1:1 BuOH-MeOH-HCO₂H-H₂O (solvent A)] of .alpha.-Asp-Ser (IIa). A brownish spot of about the same intensity [R_f 0.59 (A)], apparently representing the main product, gave a bright yellow color and corresponded to the aspartimide .alpha.,.beta.-Asp-Ser. The ir spectrum of the mixt. differed from that of IIa mainly in the carbonyl region. The absorption band was now split into well-sepd. peaks at 5.80 and 6.1 .mu., as in the case of succinimide. .alpha.-Bzl Z-glutamate (3.7 g.), 3.6 g. I, 1.4 ml. Et₃N, and 2.1 g. DCCI in 50 ml. 1:1 THF-MeCN treated like II, the soln. filtered and evapd. in vacuo, the residue dissolved in CH₂Cl₂, and the soln. filtered, washed successively with M HCl, H₂O, satd. aq. NaHCO₃, and H₂O, dried, and evapd. gave 4.1 g. Z-.gamma.-Glu-Ser-(Bzl)₂, m. 118.degree. (EtOAc-Et₂O), [.alpha.]_{27D} 1.2.degree. (c 4.0, AcOH). From 8.0 g. oily Z-isoleucine was prepd. as above 9.8 g. Z-Ile-Ser-Bzl, m. 175.degree. (EtOAc), [.alpha.]_{25D} -17.9.degree. (c 6.1, AcOH). From 8.3 g. N.alpha., N.epsilon.-Z₂-lysine was prepd. as above 9.4 g. N.alpha., N.epsilon.-Z₂-Lys-Ser-Bzl (III); when crystd. from EtOAc-ligroine, III m. 144.degree.; when crystd. from EtOAc-Et₂O, III frequently m. 122.degree., solidified, and then m. 144.degree.; the optical rotations of the 2 forms were identical, [.alpha.]_{26D} -4.8.degree. (c 6.1, AcOH), indicating dimorphism. From Z-glycine was prepd. as above Z-Gly-Ser-Bzl IIIc, m. 149.degree. (EtOAc), 4.7.degree. (c 6.1, AcOH). From 4.8 g. Z-serine and 7.0 g. alanine Bzl ester p-toluenesulfonate (IV) was prepd. 12.7 g. Z-Ser-Ala-Bzl, m. 117.degree. (EtOAc-Et₂O), [.alpha.]_{26D} -22.1.degree. (c 5.9, AcOH). The mixed anhydride, prepd. at -5.degree. from 3.7 g. .gamma.-Bzl Z-glutamate (V), 1.4 ml. Et₃N, and 0.95 ml. ClCO₂Et in 40 ml. dry 1:1 dioxane-THF, was kept 15 min. at -5.degree., a cold (-5.degree.) soln. of 2.1 g. serine in 5.0 ml. 4M NaOH added, the mixt. shaken 30 min. at room temp., the org. solvents removed in vacuo, the residual alk. soln. acidified with 3M HCl, and the product isolated with Et₂O to give 2.4 g. Z-.alpha.-Glu-Ser-.gamma.-Bzl, m. 62-4.degree. (EtOAc-ligroine); 88% dicyclohexylammonium salt (VI) m. 180.degree.. Free dipeptides (10 millimoles), 10 ml. PhCH₂OH, 11 millimoles p-MeC₆H₄SO₃H.H₂O (VII.H₂O), and 30 ml. C₆H₆ refluxed 1.5-2.0 hrs. in a Dean-Stark app., and 30 ml. C₆H₆ added gave 85-90% p-toluenesulfonate salts (isoPrOH-Et₂O) of the following peptide esters [ester, m.p., R_f in 4:1:1 BuOH-AcOH-H₂O (solvent B), and [.alpha.]_{26D} (c, AcOH) given]: Ser-Gly-Bzl (VIII), 176.degree., 0.55, 8.2.degree. (1.7); Ser-Ala-Bzl (IX), 130.degree., 0.69, -14.3.degree.

(5.5); Ser-Glu(Bzl)₂ (X), 72.degree., 0.86, -4.0.degree. (5.7). Z-Ser-Gly-Bzl (3.9 g.) refluxed 2 hrs. with 3.8 g. VII.H₂O in 35 ml. C₆H₆ contg. 5 ml. BzIOH also gave VIII p-toluenesulfonate (XI), m. 176.degree., Rf 0.55 (B), 61% yield. To 4.2 g. XI in 30 ml. MeCN and 30 ml. THF contg. 1.4 ml. Et₃N was added 3.6 g. Ia and the soln. cooled to -15.degree., treated with 2.1 g. DCCI, shaken 10 hrs. at 4.degree. and then 4 hrs. at room temp., and worked up like II to give 4.3 g. Z-.alpha.-Asp-Ser-Gly-(Bzl)₂ (XII), m. 120.degree. (EtOAc), [.alpha.]_{26D} -8.9.degree. (c 1.8, dry C₅H₅N), -8.0.degree. (c 6.0, AcOH); material m. 105-15.degree. with higher optical rotation ([.alpha.]_{26D} -12.9 to -19.4.degree.) was obtained in cases where a slight excess (10%) of Et₃N was used in the condensation step or when the reaction product was washed extensively with aq. NaHCO₃. The optical rotation of XII (c 2% in 70:30 C₅H₅N-H₂O) increased in 10 hrs. to a value of [.alpha.]_{27D} -23.3 .+- 1.0.degree. and decreased thereafter slowly. From 1.4 g. Ia, 1.7 g. IX p-toluenesulfonate, 0.66 ml. Et₃N, and 0.85 g. DCCI in 50 ml. MeCN-THF was prepd., like XII, 1.9 g. Z-.alpha.-Asp-Ser-Ala-(Bzl)₂ (XIII), m. 131.degree., [.alpha.]_{26D} -17.1.degree. (c 6.0, AcOH), -20.2.degree. (c 2.0, C₅H₅N). The optical rotation of XIII reached a value of [.alpha.]_{27D} -36.5.degree. within 3 hrs. at 27.degree. in 70:30 C₅H₅N-H₂O. A sample of XIII, prepd. by the isolation procedure involving washing with aq. NaHCO₃, m. 116-18.degree., [.alpha.]_{26.5D} -21.2.degree. (c 5.9, AcOH), indicating some aspartimide formation. Strong evidence for this was obtained by phosphorylation of 1.8 g. of this material, followed by hydrogenolysis (cf. below), giving 1.05 g. product, which was resolved by anion exchange chromatography into 2 peptides with the compn. of Asp-SerP-Ala.(H₂O)_n. The main product (fraction D, 0.53 g.) appeared at a place in the chromatogram normal for .alpha.-aspartyl-O-phosphorylserines, and gave the normal purple ninhydrin reaction on paper chromatograms Rf 0.38 (A). The optical rotation was [.alpha.]_{26D} -17.3.degree. (c 4.0, M HCl), and the analyses were correct for .alpha.-Asp-SerP-Ala.2H₂O (XIV.2H₂O). The minor component (fraction C, 166 mg.) appeared earlier in the chromatogram, indicating that it was less acidic than XIV. After isolation, it gave 1 bright yellow spot [Rf 0.44 (A)] with ninhydrin; after a year at room temp., 1 or 2 new components appeared [Rf 0.38-0.40 (A), violet color], indicating appreciable decompn. The carbonyl ir absorption band of fraction C had 2 well-sepd. peaks at 5.77 and 6.0 .mu., indicating a succinimide ring; the carbonyl ir absorption band of fraction D was broad and had the peak at 6.05 .mu.. The high optical rotation of fraction C [[.alpha.]_{26D} -40.3.degree. (c 3.8, M HCl)] was a further indication that it was N-aspartimido-O-phosphorylserylalanine-H₂O. The decompn. observed above was possibly caused by the water of hydration in opening the succinimide ring, forming a mixt. of XIV and its .beta.-isomer. Alk. phosphatase from swine kidney liberated 1 equiv. of inorg. phosphate from both fractions C and D, and at about the same rate. From 1.8 g. Ia, 2.94 g. X p-toluenesulfonate, 0.7 ml. Et₃N, and 1.1 g. DCCI was prepd. like XII 3.0 g. Z-.alpha.-Asp-Ser-Glu-(Bzl)₃ ester, m. 80-2.degree. (EtOAc-ligroine), [.alpha.]_{27D} -15.2.degree. (c 1.8, dry C₅H₅N), [.alpha.]_{26D} -15.2.degree. (c 5.9, AcOH); the optical rotation of this compd. reached a value of [.alpha.]_{27D} -32.0.degree. within 3.5 hrs. in 70:30 C₅H₅N-H₂O. From 2.8 g. V dicyclohexylamine salt (XV), 2.1 g. XI, and 1.1 g. DCCI in 80 ml. THF-MeCN was prepd. like XII 2.0 g. Z-.alpha.-Glu-Ser-Gly di-Bzl ester, m. 121.degree. (EtOAc), [.alpha.]_{26D} -7.7.degree. (c 3.3, AcOH). From 2.76 g. XV, 2.2 g. IX p-toluenesulfonate, and 1.1 g. DCCI was prepd. like XII 2.4 g. Z-.alpha.-Glu-Ser-Ala(Bzl)₂ (XVI), m. 104.degree. (EtOAc-Et₂O), [.alpha.]_{26D} -16.4.degree. (c 6.1, AcOH). VI (3.2 g.), 1.8 g. IV, and 1.1 g. DCCI in 50 ml. 1:1 THF-MeCN shaken 4 hrs. at 4.degree. and 18 hrs. at

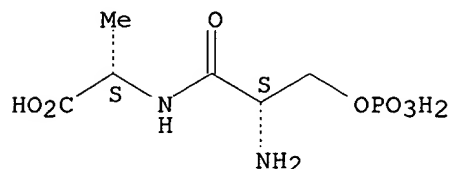
room temp., the soln. filtered and evapd., the residue treated with 150 ml. EtOAc, and the soln. filtered, washed successively with H₂O, 2M HCl, H₂O, and aq. NaHCO₃, dried, and evapd. gave 2.9 g. XVI, m. 96-8.degree. (EtOAc-Et₂O), (.alpha.)26.5D -11.2.degree. (c 6.0, AcOH). From 6.4 g. Z-Leu-Gly, 7.0 g. I, 2.8 ml. Et₃N, and 4.2 g. DCCI in 100 ml. 1:1 THF-MeCN was prepd. 8.0 g. Z-Leu-Gly-Ser-Bzl, m. 128.degree. (EtOAc-Et₂O), [.alpha.]26D -2.3.degree. (c 6.5, AcOH). The protected peptide (5 millimoles hydrogenated in tert-BuOH-H₂O over 0.5 g. 10% Pd-C (1 equiv. HCl was added in the case of the histidine and 2 equivs. in the case of the lysine peptides), when no more H was consumed (after 0.5-2.0 hrs.) the catalyst filtered off and washed with H₂O, the combined solns. evapd. in vacuo, and the residue crystd. from H₂O-EtOH gave corresponding free peptide; all free peptides gave 1 spot on chromatograms in all solvent systems tested and had the normal purple ninhydrin color; they were all tested by total hydrolysis by leucine aminopeptidase or by 6M HCl; the hydrolyzates were also analyzed by paper chromatography on Whatman No. 1 paper with solvent B, 65:35 C₅H₅-H₂O, or 10:10:5:2 BuOH-Me₂CO-H₂O-C₅H₅N. Peptide, Rf(B), Rf(A), [.alpha.]D, Temp., c in M HCl; Ser-Gly, 0.12, 0.60, 39.9.degree., 26.degree., 1.2; Ser-Ala, 0.25, 0.66, -29.5.degree., 27.degree., 1.2; Ser-Glu, 0.15, 0.62, -11.4.degree., 26.5.degree., 4.2; Ser-His, 0.03, 0.44, 14.0.degree., 26.degree., 3.9; Ile-Ser, 0.54, 0.72, 30.6.degree., 26.degree., 4.2; Asp-Ser, 0.07, 0.50, 24.0.degree., 27.degree., 1.7; Glu-Ser, 0.08, 0.46, 19.5.degree., 26.5.degree., 4.0; Lys-Ser, 0.06, 0.42, 27.9.degree., 27.degree., 3.8; Asp-Ser-Gly, 0.06, 0.47, -5.0.degree., 26.5.degree., 4.1; , , , , -8.2.degree., (c 3.2, H₂O); Asp-Ser-Ala, 0.08, 0.58, -34.0.degree., 26.degree., 1.6; Asp-Ser-Glu, 0.07, 0.48, -19.5.degree., 26.5.degree., 4.0; , , , , -15.5.degree., (c 3.2, H₂O); Glu-Ser-Gly, 0.09, 0.45, 8.3.degree., 27.degree., 4.4; Glu-Ser-Ala, 0.14, 0.58, 16.2.degree., 26.5.degree., 1.6; Leu-Gly-Ser, 0.34, 0.75, 36.5.degree., 26.degree., 3.9. The peptides listed in the first table were prepd. The Z-protected peptide benzyl ester (5 millimoles) was dissolved in 10 ml. dry (over BaO) C₅H₅N and the soln. cooled to just above the f.p. Dibenzylphosphoryl chloride (XVII), freshly prepd. from 1.9 g. dibenzyl phosphite and N-chlorosuccinimide, was added. The mixt. was shaken and kept overnight at 4.degree.. Cold EtOAc (75 ml.) and 75 ml. cold H₂O were added, and the upper phase washed successively with cold H₂O, M H₂SO₄, H₂O, satd. aq. NaHCO₃, and H₂O, dried, and evapd. in vacuo to give phosphate triesters O,O-dibenzylphosphorylated protected serine peptides (XVIII) as solids or semisolids. When XVII was used in excess, it was possible, after long reaction times, to detect the formation of pyrophosphate and triphosphate analogs of the O-phosphorylated peptides. XVIII were hydrogenolyzed, in most cases, without further purification, although some of them may be crystd. Thus, 87% Z-glycyl(O,O-dibenzylphosphoryl)serine Bzl ester (XIX) was obtained, m. 79-80.degree. (EtOAc), Rf 0.42 [thin layer chromatography (TLC) on silica gel G with 60:40:1 ligroine-THF-AcOH (solvent C)]. Since IIIa had Rf 0.25 (C), chromatography on silicic acid may be an alternative purification procedure for XVIII and XIX. A third method for purification is the monodebenzylation procedure of Zervas and Dilaris (CA 50, 6350e). Thus, 6.5 g. XIX in 20 ml. dry Me₂CO refluxed 45 min. with 1.5 g. dry NaI gave 4.3 g. diester Na salt, C₂₇H₂₈N₂O₉PNa, m. 178.degree., Rf 0.0 (TLC with C), which (3.5 g.) dissolved in 25 ml. H₂O and treated with 7 ml. M HCl gave 3.3 g. IIIa O-Bzl H phosphate (XX), m. 129.degree. (EtOAc-Et₂O), Rf 0.0 (TLC with C). XX (1.5 g.) treated with 0.28 g. DCCI in 15 ml. dry CH₂Cl₂ and after 20 min. at room temp. the soln. filtered and evapd. in vacuo gave 1.5 g. semisolid residue (XXI), Rf 0.63 (TLC with C). XXI was transformed to XX, m. 127-8.degree., Rf 0 (TLC with C), when left several

days in an open vessel. These facts, together with the method of synthesis, indicate XXI to be $[\text{PhCH}_2\text{O}_2\text{CNHCH}_2\text{CONHCH}(\text{CO}_2\text{CH}_2\text{Ph})\text{CH}_2\text{OP}(\text{O})(\text{OCH}_2\text{Ph})]_2$. XVIII were hydrogenolyzed over 10% Pd-C (0.3-0.5 g./g. XVIII) in tert-BuOH-H₂O as described for the prepn. of the free peptides, 1-4 hrs. shaking in a H atm. being required. The soln. filtered and evapd. in vacuo gave the phosphorylated peptide, some of which crystd. from H₂O-EtOH (cf. run-on table). However, the majority of the phosphopeptides were purified by chromatography on a column of anion exchange resin, Dowex 1-X2 (formate form), as previously described (Avaeva, et al., CA 60, 12099c). The freeze-dried phosphopeptide fractions were mostly, according to analysis, in the form of monopyridinium salts. They were, therefore, triturated in H₂O with a small amt. cation-exchange resin, Dowex 50 (H⁺ form), and the soln. filtered and again freeze-dried. The following paper chromatographically pure peptides were obtained in this way in 51-81% yields: [peptide, Rf(B), Rf(A); [.alpha.]D, temp., and c (M HCl)] given: SerP-Ala (H₂O-EtOH), 0.09, 0.48; -16.5.degree., 26.degree., 4.2: SerP-His, 0.0, 0.27; 9.4.degree., 27.degree., 1.9: Asp-SerP, 0.03, 0.32; 21.6.degree., 27.degree., 4.2: Glu-SerP, 0.03, 0.29; 24.7.degree., 27.degree., 4.0: Ile-SerP (H₂O-EtOH), 0.18, 0.62; 30.5.degree., 27.degree., 4.0: Lys-SerP, 0.03, 0.30; 25.2.degree., 25.degree., 4.0: Asp-SerP-Gly, 0.02, 0.30; -4.3.degree., 26.degree., 3.9: Asp-SerP-Ala 0.03, 0.38; -17.3.degree., 26.degree., 4.0: Asp-SerP-Glu, 0.02, 0.33; -11.2.degree., 26.degree., 4.2: Glu-SerP-Gly, 0.02, 0.35; 11.5.degree., 27.degree., 4.0: Glu-SerP-Ala, 0.04, 0.40; -4.5.degree., 25.degree., 4.3: Leu-Gly-SerP (H₂O-EtOH), 0.24, 0.62; 40.6.degree., 27.degree., 4.1. Lys-SerP and SerP-His were C₅H₅N-free after the first freeze-drying. SerP-His was obtained by phosphorylation of N-(N-Z-Ser)-Nim-Bzl-His-Bzl. The hydrogenolysis required 12 hrs. for complete removal of the Nim-Bzl group. The procedure was used without difficulties also in the prepn. of SerP-Gly-2-14C and Gly-SerP-3-14C from glycine-2-14C and serine-3-14 C. O-Phosphoryl-DL-serine (XXII) (6.8 g.) added to a soln. (prepd. at -10.degree.) of 26 ml. SOCl₂ in 100 ml. MeOH (XXII dissolved completely within 1 hr.) and the soln. evapd. in vacuo gave 7.3 g. XXII Me ester (XXIII), m. 198.degree. (decompn.) (H₂O-MeOH), Rf 0.19 (B), 0.55 (in 80:20 PhOH-H₂O, identical (mixed m.p., Rf values, and ir spectrum) with XXIII prepd. (CA 53, 22157f) by hydrogenolysis of N-Z-(O,O-diphenylphosphoryl)-DL-serine Me ester. O-Phosphorylserine (4.7 g.) treated similarly gave 3.7 g. corresponding optically active Me ester, m. 167.degree. (decompn.), [.alpha.]_{26D} 12.0.degree. (c 4.3, M HCl), Rf 0.19 (B). Threonine (29 g.; [.alpha.]_{26D} -14.4.degree. (c 2.1, 5N HCl)) treated with chlorophosphoric acid as described for serine (CA 53, 22157f) and after hydrolysis EtOH and Et₂O added gave 22.8 g. O-phosphorylthreonine (XXIV), m. 189.degree. (decompn.) (H₂O-EtOH), [.alpha.]_{27D} -7.9.degree. (c 2.5, H₂O), Rf 0.11 (B), 0.46 (A), 0.03 (in 35:35:30 C₅H₅N-iso-AmOH-C₂O), the m.p. and [.alpha.]_D being in accord with those of XXIV isolated by Verdier (CA 47, 5971b) from an acid hydrolyzate of bovine casein. The acid hydrolyzate (20 hrs., 110.degree., sealed tubes) of synthetic XXIV (38.7 mg. in 2200.1 mg. 5M HCl) had negligible optical rotation, [.alpha.]_{26D} -1.5.degree.. Paper chromatographic analysis of the hydrolyzate indicated threonine to be the main product present. In addn., a small amt. unchanged XXIV was present. Little loss of optical activity was observed when threonine was heated similarly in 5M HCl (41.4 mg. in 2167.0 mg.; [.alpha.]_{26D} -13.2.degree. (c 1.7)). When H₃PO₄ (24.5 mg. 85% H₃PO₄ was present during the hydrolysis of threonine (40.0 mg. in 1987.6 mg. 5M HCl), the rotation became [.alpha.]_{26D} -12.8.degree. (c 1.8).

IT 6403-04-9, Alanine, N-L-seryl-, dihydrogen phosphate (ester), L- (prepn. of)

RN 6403-04-9 HCAPLUS
 CN Alanine, N-L-seryl-, dihydrogen phosphate (ester), L- (8CI) (CA INDEX NAME)

Absolute stereochemistry.



L19 ANSWER 18 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1966:28766 HCAPLUS

DOCUMENT NUMBER: 64:28766

ORIGINAL REFERENCE NO.: 64:5369f-g

TITLE: Hydrolysis of phosphopeptides. III. The action of alkaline phosphatase preparations from kidney, bone, and yeast on O-phosphorylated model compounds

AUTHOR(S): Csopak, Hedvig; Folsch, Georg; Strid, Lars; Mellander, Olof

CORPORATE SOURCE: Univ. Goteborg, Swed.

SOURCE: Acta Chem. Scand. (1965), 19(7), 1575-82

DOCUMENT TYPE: Journal

LANGUAGE: English

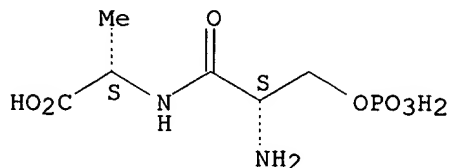
AB cf. preceding abstr. Swine kidney, calf bone, and yeast phosphatase enzymes were prepd. and the rate of PO₄³⁻ liberation from 30 phosphopeptides was detd. The enzymes were not purified sufficiently to exclude the possibility of the presence of enzymes other than phosphatases. Position of the PO₄³⁻ affected the rate of hydrolysis. It was faster when in the N-terminal position than when in the reversed sequences. The ext. from the kidney tissue was more active than that from bone tissue. The ext. from yeast caused hydrolysis in 10 peptides. 21 references.

IT **6403-04-9**, Alanine, N-L-seryl-, dihydrogen phosphate (ester), L-
6665-27-6, Leucine, N-L-seryl-, dihydrogen phosphate (ester), L-
6665-42-5, Glycine, N-L-seryl-, dihydrogen phosphate (ester)
 (hydrolysis by phosphatase)

RN 6403-04-9 HCAPLUS

CN Alanine, N-L-seryl-, dihydrogen phosphate (ester), L- (8CI) (CA INDEX NAME)

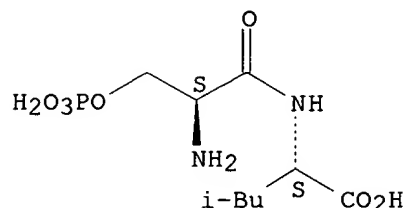
Absolute stereochemistry.



RN 6665-27-6 HCAPLUS

CN L-Leucine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

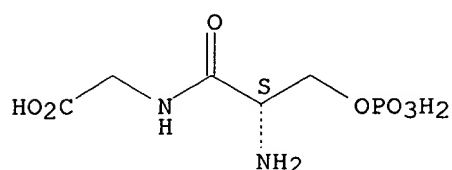
Absolute stereochemistry.



RN 6665-42-5 HCAPLUS

CN Glycine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



L19 ANSWER 19 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1966:28765 HCAPLUS

DOCUMENT NUMBER: 64:28765

ORIGINAL REFERENCE NO.: 64:5369c-f

TITLE: Hydrolysis of phosphopeptides. II. Leucine
aminopeptidase hydrolysis of free and O-phosphorylated
serine peptides

AUTHOR(S): Folsch, Georg; Strid, Lars; Mellander, Olof

CORPORATE SOURCE: Univ. Goteborg, Swed.

SOURCE: Acta Chem. Scand. (1965), 19(7), 1566-74

DOCUMENT TYPE: Journal

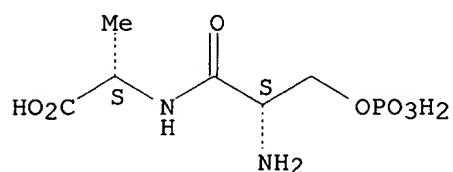
LANGUAGE: English

AB cf. CA 56, 6083i; following abstr. The relative rate of hydrolysis by leucine aminopeptidase (I) of substrates consisting of phosphate-free peptides (II), O-phosphorylated (III), O-monophenylphosphorylated (IV), and O-pyrophosphorylated serine peptides (V) was detd. The enzyme hydrolyzed peptides having N-terminal O-phosphorylserine only about 0.4% as fast as the corresponding nonphosphorylated peptides. Dipeptides with O-phosphorylserine in the C-terminal position were hydrolyzed approx. 1% as fast as were the corresponding serine peptides with a free hydroxyl group. The hydrolysis rate was directly proportional to the distance of the PO43- group from the free .alpha.-amino end. A pyrophosphoryl group retarded hydrolysis even more than a simple PO43- group. The attachment of a hydrophobic benzene ring to the PO43- group increased the rate of hydrolysis to the same or even higher value than that of the nonphosphorylated analogs. Percent hydrolysis in 20 min., using 0.1% of I, varied from 20.0 to 0.01 for the 15 II tested, 7.4 to 0.001 for 11 III tested, 0.58 to 0.04 for the 2 IV, and 0.055 to 0.003 for the 2 V substrates. Compds. tested for hydrolysis from the highest to the lowest rate in each group were: II, L-Leu-Gly, L-Leu-L-Ser, L-Leu-Gly-L-Ser, L-Ser-L-Leu, L-Ile-L-Ser, Gly-L-Leu, .alpha.-L-Glu-L-Ser-L-Ala,

L-Lys-L-Ser, L-Ser-Gly, DL-Ser-Gly, L-Ser-L-Ala, Gly-DL-Ser, .gamma.-L-Glu-L-Ser, Gly-Gly, and .alpha.-L-Glu-L-Ser; group III, L-Leu-Gly-SerP, L-Leu-SerP, L-SerP-L-Leu, L-Ile-L-SerP, L-Lys-L-SerP, L-SerP-L-Ala, DL-SerP-Gly, Gly-DL-SerP, L-SerP-L-SerP, and .gamma.-L-Glu-L-SerP; group IV DL-Ser.vphi.P-Gly, and Gly-DL-Ser.vphi.P; and group V L-Leu-L-SerPP, and L-SerPP-L-Leu. The above P represents O-phosphorylated, the .vphi.P O-monophenylphosphorylated, and PP O-pyrophosphorylated. 29 references.

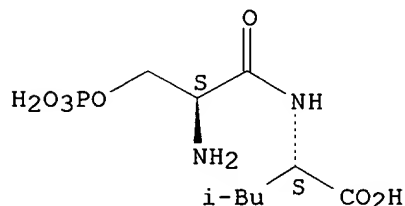
IT 6403-04-9, Alanine, N-L-seryl-, dihydrogen phosphate (ester), L-
 6665-27-6, Leucine, N-L-seryl-, dihydrogen phosphate (ester), L-
 6665-32-3, Glycine, N-DL-seryl-, dihydrogen phosphate (ester)
 (hydrolysis by leucine aminopeptidase)
 RN 6403-04-9 HCAPLUS
 CN Alanine, N-L-seryl-, dihydrogen phosphate (ester), L- (8CI) (CA INDEX NAME)

Absolute stereochemistry.

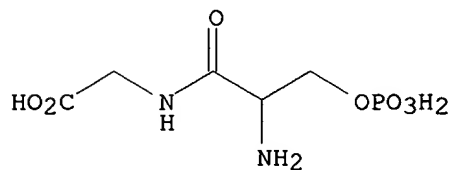


RN 6665-27-6 HCAPLUS
 CN L-Leucine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



RN 6665-32-3 HCAPLUS
 CN Glycine, N-DL-seryl-, dihydrogen phosphate (ester) (8CI) (CA INDEX NAME)



L19 ANSWER 20 OF 24 HCAPLUS COPYRIGHT 2002 ACS
 ACCESSION NUMBER: 1965:455686 HCAPLUS
 DOCUMENT NUMBER: 63:55686
 ORIGINAL REFERENCE NO.: 63:10199a-b

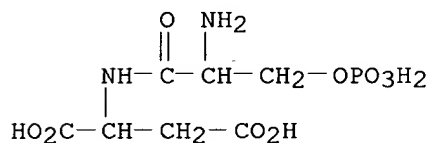
TITLE: Phosphopeptides. Synthesis and enzymic studies
 AUTHOR(S): Koehn, Paul V.; Kind, C. Albert
 CORPORATE SOURCE: Univ. of Connecticut, Storrs
 SOURCE: Arch. Biochem. Biophys. (1965), 111(3), 614-18
 DOCUMENT TYPE: Journal
 LANGUAGE: English

AB Phosphoprotein phosphatase obtained from chick embryo readily dephosphorylates casein which is characterized by sequences of O-phosphorylserine residues and by a high content of glutamic acid and aspartic acid. Dipeptides incorporating these amino acids and (SerP)3 were synthesized. The phosphopeptides, equivalent to 380 .gamma. P in acetate buffer (pH 5.8). were dephosphorylated by acid phosphatase but not by phosphoprotein phosphatase. Possible min. structural features of a peptide required for phosphoprotein phosphatase activity are discussed.

IT **3785-84-0**, Aspartic acid, N-seryl-, dihydrogen phosphate (ester) (prepn. and diphosphorylation of)

RN 3785-84-0 HCAPLUS

CN Aspartic acid, N-seryl-, dihydrogen phosphate (ester) (8CI) (CA INDEX NAME)



L19 ANSWER 21 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1963:60714 HCAPLUS
 DOCUMENT NUMBER: 58:60714
 ORIGINAL REFERENCE NO.: 58:10423a-c
 TITLE: Calcium, magnesium, and manganese(II) complexes of some O-phosphorylated peptides
 AUTHOR(S): Osterberg, Ragnar
 CORPORATE SOURCE: Univ. Goteborg, Swed.
 SOURCE: Acta Chem. Scand. (1962), 16, 2434-51
 DOCUMENT TYPE: Journal
 LANGUAGE: English

AB Complex formation between Ca, Mg, and Mn(II) ions and O-phosphorylated ethanolamine, seryllysine, serylglycine, glycylserine, glycylserylglycine, and serylglutamic acid was studied by pH titration at 25.degree. in a medium contg. const. Cl- concn. Complexes of the type MpHqA were formed where $p:q = 1:1$ and $1:0$ for the first two ligands, $p:q = 1:1$, $1:0$, and $2:0$ for the next three ligands (although $p:q = 2:0$ was not measurable for $\text{M} = \text{Ca}^{++}$), and $p:q = 1:2$, $1:1$, $1:0$, $2:1$, and $2:0$ for the last ligand. Stability consts. for all indicated complexes were tabulated and the coordination sites discussed. The observed differences between Ca^{++} and Mg^{++} in complexing ability was correlated to ionic size. The definitely greater tendency of Mg^{++} and Mn^{++} ions to dinuclear complex formation compared with Ca^{++} was discussed in relation to the specificity of enzymes for these ions.

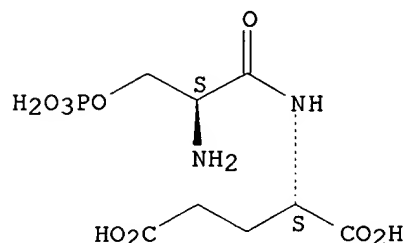
IT **93115-01-6**, Hydrogen [N-L-seryl-L-glutamic acid phosphato(4-)]magnesate **93168-28-6**, Hydrogen [N-L-seryl-L-glutamic acid phosphato(4-)]calciate **93285-19-9**, Hydrogen [N-DL-serylglycine phosphato(3-)]calciate **93409-37-1**,

Hydrogen [N-L-seryl-L-glutamic acid phosphato(3-)]aquocalciate
93440-75-6, Manganese, [(N-L-seryl-L-glutamic acid
 phosphato(4-)]tetraaquodi- **93481-54-0**, Calcium,
 [N-L-seryl-L-glutamic acid phosphato(4-)] tetraaquodi- **93784-32-8**
 , Magnesium, [N-L-seryl-L-glutamic acid phosphato(4-)]tetraaquodi-
 (prepn. of)

RN 93115-01-6 HCAPLUS

CN Hydrogen [N-L-seryl-L-glutamic acid phosphato(4-)]magnesate (7CI) (CA
 INDEX NAME)

Absolute stereochemistry.

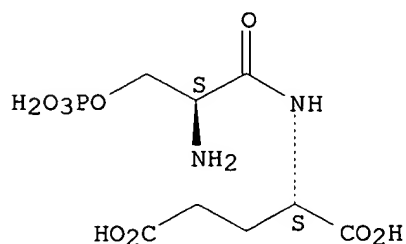


● Mg

RN 93168-28-6 HCAPLUS

CN Hydrogen [N-L-seryl-L-glutamic acid phosphato(4-)]calciate (7CI) (CA
 INDEX NAME)

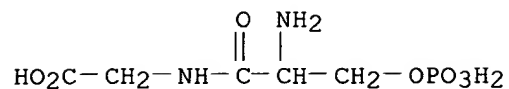
Absolute stereochemistry.



● Ca

RN 93285-19-9 HCAPLUS

CN Hydrogen [N-DL-serylglycine phosphato(3-)]calciate (7CI) (CA INDEX NAME)

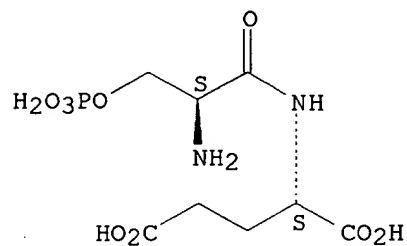


● Ca

RN 93409-37-1 HCAPLUS

CN Hydrogen [N-L-seryl-L-glutamic acid phosphato(3-)]aquocalciate (7CI) (CA INDEX NAME)

Absolute stereochemistry.



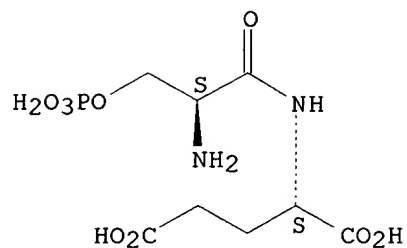
● Ca

● H₂O

RN 93440-75-6 HCAPLUS

CN Manganese, [N-L-seryl-L-glutamic acid phosphato(4-)]tetraaquodi- (7CI) (CA INDEX NAME)

Absolute stereochemistry.



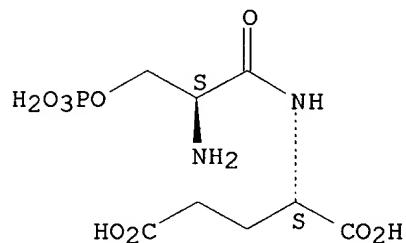
●2 Mn(II)

●4 H₂O

RN 93481-54-0 HCAPLUS

CN Calcium, [N-L-seryl-L-glutamic acid phosphato(4-)]tetraaquodi- (7CI) (CA INDEX NAME)

Absolute stereochemistry.



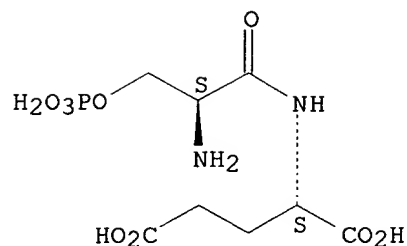
●2 Ca

●4 H₂O

RN 93784-32-8 HCAPLUS

CN Magnesium, [N-L-seryl-L-glutamic acid phosphato(4-)]tetraaquodi- (7CI) (CA INDEX NAME)

Absolute stereochemistry.



●2 Mg

●4 H₂O

L19 ANSWER 22 OF 24 HCAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1963:60713 HCAPLUS

DOCUMENT NUMBER: 58:60713

ORIGINAL REFERENCE NO.: 58:10422h,10423a

TITLE: Biosynthesis of amino sugar in hyaline cartilage

AUTHOR(S): Seno, Nobuko

CORPORATE SOURCE: Ochanomizu Univ., Tokyo

SOURCE: Seikagaku (1962), 34, 629-31

DOCUMENT TYPE: Journal

LANGUAGE: Unavailable

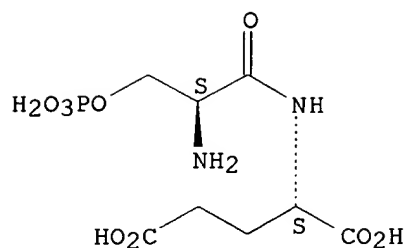
AB An enzyme to synthesize amino sugar from glucose 1-phosphate and glutamine was prepd. from hyaline cartilage tissues. The optimum pH of the reaction was 7.4, the same as rat liver enzyme. An (NH₄)₂SO₄ pptg. fraction at higher than 2.3M showed the highest enzyme activity, whereas a fraction pptg. at 1.7-2.3M satn. was the most active for rat liver enzyme. Enzyme activity (amino sugar formation micromoles/mg. enzyme protein N) of hyaline cartilage was about 2.5 times higher than that of rat liver.

IT **93168-28-6**, Hydrogen [N-L-seryl-L-glutamic acid phosphato(4-)]calciate **93285-19-9**, Hydrogen [N-DL-serylglycine phosphato(3-)]calciate **93409-37-1**, Hydrogen [N-L-seryl-L-glutamic acid phosphato(3-)]aquocalciate (prepn. of)

RN 93168-28-6 HCAPLUS

CN Hydrogen [N-L-seryl-L-glutamic acid phosphato(4-)]calciate (7CI) (CA INDEX NAME)

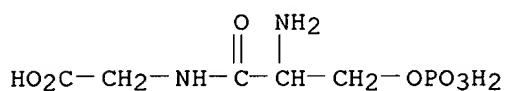
Absolute stereochemistry.



● Ca

RN 93285-19-9 HCAPLUS

CN Hydrogen [N-DL-serylglycine phosphato(3-)]calcite (7CI) (CA INDEX NAME)

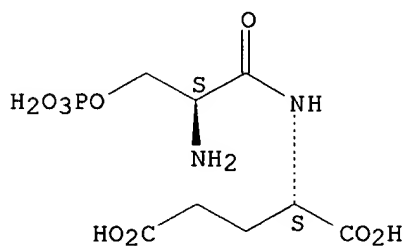


● Ca

RN 93409-37-1 HCAPLUS

CN Hydrogen [N-L-seryl-L-glutamic acid phosphato(3-)]aquocalcite (7CI) (CA INDEX NAME)

Absolute stereochemistry.



● Ca

● H2O

L19 ANSWER 23 OF 24 HCAPLUS COPYRIGHT 2002 ACS
 ACCESSION NUMBER: 1962:42074 HCAPLUS

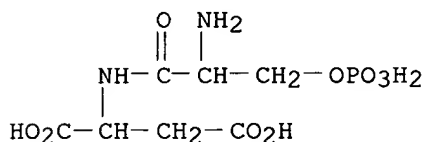
DOCUMENT NUMBER: 56:42074
 ORIGINAL REFERENCE NO.: 56:7975a-c
 TITLE: Separation of O-phosphorylated amino acids and peptides on anion exchange resin
 AUTHOR(S): Strid, Lars
 CORPORATE SOURCE: Univ. Goteborg, Swed.
 SOURCE: Acta Chem. Scand. (1959), 13, 1787-90
 DOCUMENT TYPE: Journal
 LANGUAGE: English

AB The following compds. were sepd. on a 1.6 .times. 50-cm. anion-exchange column contg. Dowex 1-X2(Cl-), 200-400 mesh: O-phosphorylserine, O-phosphorylthreonine, glycyl-(O-phosphoryl)serylglycine, glycyl-(O-phosphoryl)serine, leucyl-(O-phosphoryl)serine, O-phosphorylseryl-(O-phosphoryl)serine, and O-phosphorylserylglycine, -leucine, -aspartic acid, and -glutamic acid. By slowing passing 2M HCO₂Na, pH 5.5, through the column until the effluent gave a neg. Cl- test, the resin was converted to the formate. Phosphorylated amino acids and phosphopeptides contg. neutral amino acids are eluted within a narrow region, the order of elution agreeing with the pK value of the CO₂H group.

IT **3785-84-0**, Aspartic acid, N-seryl-, dihydrogen phosphate (ester) (chromatography of)

RN 3785-84-0 HCAPLUS

CN Aspartic acid, N-seryl-, dihydrogen phosphate (ester) (8CI) (CA INDEX NAME)



L19 ANSWER 24 OF 24 HCAPLUS COPYRIGHT 2002 ACS

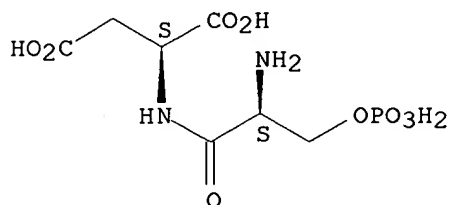
ACCESSION NUMBER: 1962:38740 HCAPLUS
 DOCUMENT NUMBER: 56:38740
 ORIGINAL REFERENCE NO.: 56:7418e-i
 TITLE: Synthesis of phosphopeptides. II. O-Phosphorylated dipeptides of L-serine
 AUTHOR(S): Folsch, George
 CORPORATE SOURCE: Univ. Gotheeburg, Swed.
 SOURCE: Acta Chem. Scand. (1959), 13, 1407-21
 DOCUMENT TYPE: Journal
 LANGUAGE: English

AB cf. CA 53, 17919h.-Diphenyl- and dibenzylphosphoryl chlorides phosphorylated 6-(N-carbobenzoxo)dipeptide benzyl esters of L-serine with amino acid sequences found in phosphoproteins; catalytic hydrogenolysis of the dibenzylphosphoryl derivs. gave pure phosphopeptides, while the diphenylphosphoryl analogs frequently gave partially hydrogenated derivs. Compds. prepd. include the following L,L-N-carbobenzoxo dipeptide benzyl esters (compd., % yield, and m.p. given, Z = PhCH₂OCO): Z-ser-glu-OCH₂Ph, 78, 125.degree.; Z-ser-asp-OCH₂Ph, 89, 128-9.degree.; Z-ser-leu-OCH₂Ph, 90, 83-4.degree.; Z-ser-ser-OCH₂Ph, 71, 138-9.degree.; Z-glu-ser-OCH₂Ph, 76, 127;; and Z-leu-ser-OCH₂Ph, 86, 122-3.degree.. The following dipeptides of L-serine [compd., % yield, and [.alpha.]₂₁D (N HCl, 1 l) given]: H-ser-glu-OH, 82, -10.8.degree. (c 4.4); H-ser-asp-OH, 78,

1.5.degree. (c4.0); H-ser-leu-OH, 79, -18.8.degree. (c 2.4); H-ser-ser-OH, 74, 14.8.degree. (c 6.4); H-glu-ser-OH, 80, 29.0.degree. (c 5.1); and H-leu-ser-OH, 74, 27.8.degree. (c 6.9). The following dipeptides of O-phosphoryl-L-serine [compd., % yield, and [.alpha.]21D (N HCl, 1 l) given; ser P = NHCH[CH2OPO(OH)2]CO]; H-ser-P-glu-OH (I), 53, -9.5.degree. (c 4.2); H-ser-P-asp-OH, 62, -2.2.degree. (c 2.7); H-ser-P-leu-OH, 64, -16.0.degree. (c 4.0); H-ser-P-ser-OH, 57, 8.2 (c 4.7); H-glu-ser-P-OH, 54, 23.3.degree. (c 4.4); and H-leu-ser-P-OH, 62, 24.5.degree. (c 7.0). The following dibenzyl, di-Ph, and Ph phosphorylated derivs. (compd., % yield, and m.p. given): Z-ser[OPO(OCH2Ph)2]-gly-OCH2Ph, 75, 104-5.degree.; Z-gly-ser[OPO(OCH2Ph)2]-OCH2Ph, 76, 81-2.degree.; Z-ser[OPO(OPh)2]-asp-(OCH2Ph)2, 93, 62-3.degree.; Z-glu(OCH2Ph)ser[OPO(OPh)2]-OCH2Ph, 90, 75-6.degree.; H-ser[OPO(OPh)2]-glu(OH)2, 48, 182-5.degree. (decompn.); H-ser[OPO(OPh)OH]-glu(OH)2, 21, 174-8.degree. (decompn.); H-ser[OPO(OPh)OH]-asp(OH)2, 28, 167-71.degree. (decompn.); and H-ser[OPO(OPh)-OH]-leu-OH, 11, 202-4.degree. (decompn.). Also prepd. were H-ser[OPO(OPr)2OCH2Ph], m. 130-1.degree. (decompn.), the brucine salt of I, m. 171-3.degree. (decompn.), and the PhSO3H, p-MeC6H4SO3H, and HCl salts of L-serine benzyl ester, m. 110-12.degree., 94-5.degree., and 172-4.degree., [.alpha.]21D -4.1.degree. (MeOH, c4.4, 1 l), resp.

IT 1492-20-2, Aspartic acid, N-L-seryl-, dihydrogen phosphate (ester), L- 6665-27-6, Leucine, N-L-seryl-, dihydrogen phosphate (ester), L- (prepn. of)
 RN 1492-20-2 HCAPLUS
 CN L-Aspartic acid, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.



RN 6665-27-6 HCAPLUS
 CN L-Leucine, N-(O-phosphono-L-seryl)- (9CI) (CA INDEX NAME)

Absolute stereochemistry.

